1 **11.** Diatoms in temporary rivers: importance in a global climate change context 2

3 Maria Helena Novais^{1,2}, Manuela Morais^{1,2}

4

¹ICT, Institute of Earth Sciences, University of Évora, Rua Romão Ramalho 59, Évora, Portugal

6 ²Water Laboratory, University of Évora, P.I.T.E. Rua da Barba Rala Nº 1, 7005-345 Évora,

7 Portugal

- 8 E-mail: <u>hnovais@uevora.pt</u>; <u>mmorais@uevora.pt</u>
- 9

10 Abstract

In this chapter we describe the biological aspects of diatoms in temporary rivers, focusing on their 11 12 ecology and adaptation mechanisms to the harsh environment in these aquatic ecosystems. 13 Predicted alterations in temporary rivers, according to the climate change scenarios, will most probably affect diatom composition and community structure. Given the importance of diatoms 14 15 in these ecosystems, changes in diatom taxonomical and functional diversity, physiology, or biomass will be reflected in the whole ecosystem. The adaptation mechanisms to the stressors in 16 17 temporary rivers can contribute to providing diatoms with an adaptive advantage under the predicted climate change extreme events. Furthermore, the use of diatoms as bioindicators, 18 19 including the dry phase, contributes to the conservation and management of temporary rivers. 20 However, further studies on diatoms in temporary rivers should be carried out, since their ecology 21 is still poorly known due to the lack of exhaustive and thorough studies on their taxonomy, 22 highlighted by the number of new species recently described from a few sampling sites in 23 Southern Portugal.

24

25 11.1 Introduction

Watercourses that are subject to flow cessation, include a wide range of hydrological regimes, namely, temporary or intermittent rivers/streams, that may dry up during some time along the year [11.1], and ephemeral streams, that only flow for a short period, usually after rainfall events [11.1] or after snow melt [11.2]. To describe these watercourses a diverse lexicon is applied, as temporary, intermittent, seasonally intermittent, non-perennial, episodic or ephemeral [11.3], or freezing intermittent rivers and ephemeral streams (FIRES - lotic systems that have the whole water column
periodically freezing) [11.4]. In this review, the term "temporary rivers" was selected.

34 The integration of temporary rivers in riverine ecosystems conceptual models has been 35 critically revised by [11.5], stating that the 18 models jointly describe the main drivers of 36 biogeochemical and ecological patterns and processes along the following four 37 dimensions: (i) longitudinally (upstream-downstream); (ii) laterally (channel-riparian-38 floodplain); (iii) vertically (surface water-groundwater); and (iv) temporally across local 39 and landscape scales. Nevertheless, drying represents a significant hydrological process 40 that structures river ecosystems, contributing to the specificities of temporary watercourses that are not represented in these models, namely: 41

- 42 (i) their longitudinal discontinuity, with the formation of isolated pools or
 43 disconnected flowing reaches during the dry phase, with consequences for
 44 materials transport and for the connectivity of the populations of riverine
 45 organisms [11.6];
- 46 (ii) the unidirectional lateral connectivity during part of the year (only terrestrial
 47 to aquatic ecosystems);
- the variability in the vertical connectivity between surface and subsurface (iii) 48 water throughout the year, since it can be unidirectional during the drying 49 (downwelling of surface water into the hyporheic zone, the saturated 50 subsurface zone beneath the river channel) and rewetting phases (upwelling 51 of subsurface water into the river channel). This variability in the vertical 52 53 connectivity affects the nutrient and organic matter dynamics [11.7], the vertical exchanges of gases, with litter rewetting contributing to CO2 54 55 emissions [11.8], and the hyporheic zone can act as a refuge for the microorganisms [11.9]; 56
- (iv) changes in flow, caused by the drying process should be considered across
 temporal and spatial scales [11.5]. Thus, these systems are typically
 characterized by alternation between wet and dry phases, and are archetypal
 aquatic-terrestrial ecosystems, including flowing, ponded, and dry habitats
 [11.10], as presented in Figures 11.1 and 11.2.

Temporary rivers are considered the most widespread type of flowing waters, dominatingarid, semi-arid, Mediterranean, and dry-subhumid regions, together covering almost half

- of the land surface [11.2]. Also [11.11] estimate that respectively 69, 56, 49, 42 and 34
- 65 % of stream orders 1-5 are ephemeral in COSCATs (coastal segmentation and related
- 66 catchment) below 60° N, therefore, these ecosystems are important even in temperate and
- 67 humid areas, since headwaters represent more than 70% of the channel length of most
- river networks [11.12]. Recent estimations indicate that 51-60% of the global river length
- is non-perennial [11.13], and in Southern Portugal represents >70% of the river network
- 70 (Agência Portuguesa do Ambiente, I.P.; A.R.H. Alentejo, unpublished data).
- 71 [Figure 11.1 here]
- Figure 11.1. Example of dry and ponded habitats present in temporary rivers. Lampreia stream, Southern Portugal
 (left photo); Cuncos stream (right photo), both in Guadiana Basin, Southern Portugal.
- 74

75 [Figure 11.2 here]

Figure 11.2. Temporal dynamics of a temporary river with the presence of wet and dry phases. Valverde stream
(Sado Basin), Southern Portugal.

78

Further, temporary rivers are increasing annually, with more of the land surface drying, due to the reduction and seasonal shifts in precipitation and runoff [11.2]. Temporary rivers are among the aquatic ecosystems more susceptible to being affected by climate change [11.14]. This intermittence increase (both in duration, frequency, and extension, in terms of stretches/areas subject to drying) triggered by climatic drying is aggravated by human activities, such as water abstraction and diversion, with the Anthropocenic intermittence [11.2] creating artificially intermittent reaches.

The importance of naturally temporary rivers for biodiversity support, ecosystem 86 87 processes and for goods and services provision is already acknowledged [11.2], [11.15], [11.16]. Despite this recent recognition of their value and global increasing spatial and 88 89 temporal extent, temporary rivers are still largely omitted from legislation and policies, even though the Mediterranean GIG river types recognized by the WFD [11.17], already 90 91 included "small, lowland, temporary" rivers. Consequently, these rivers are still typically poorly represented in biomonitoring programs, jeopardizing their assessment, and 92 93 impairing the adoption of adequate management measures to prevent their health degradation [11.18], [11.19]. Temporary rivers are highly vulnerable to multiple 94 95 anthropogenic pressures, namely agriculture intensification, livestock farming, 96 hydromorphological alteration, and urban wastewater discharge, and these pressures will
97 likely increase under global change [11.20]–[11.22].

Research on these ecosystems has increased over the past decades, as depicted in a search 98 99 on Scopus in January 2023, where a total of 3 072 manuscripts were published with the terms "temporary river*" OR "intermittent stream*" OR "temporary stream*" OR 100 "intermittent river*" OR "ephemeral stream" OR "non-perennial river*" OR "non-101 102 perennial stream*". The first record of a published paper on this subject was entitled "Intermittent Streams in Berkshire", written by Rupert Jones in 1885 and published in the 103 104 Geological Magazine. During the following decades, almost no interest was devoted to 105 this subject, until the 60's, when one or two papers started to be published each year, 106 increasing in the 70's and onward, until surpassing the threshold of more than 100 publications per year in 2011. Since this date, more than 100 papers were published every 107 108 year, with a maximum of 193 publications in 2020, as seen in Figure 11.3.

From these, when the search term "alga*" is included, 95 publications are retrieved, from 109 which only 56 are related to diatoms. The first published record on diatoms in temporary 110 rivers is from 1985, and until 2007 only one paper was published per year. In 2008, 2 111 papers were published, with this number increasing to 4 in 2014 and reaching a maximum 112 of 7 in 2020 and 2021, revealing a slight increase in interest in this subject, as seen in 113 Figure 11.3. These studies include taxonomical approaches, with the description of 3 new 114 taxa from temporary rivers (Punctastriata obstinata E.Morales, M.L.García & Novais; 115 116 Planothidium audax E.Morales, Novais & M.L.García; Fragilaria odeloucaensis 117 E.Morales, Novais, C.E.Wetzel, Ector & Morais); community dynamics; monitoring and ecological status assessment, detailed in the following sections. The great majority of the 118 119 studies are focused in drying rivers, whilst diatom studies in intermittent freezing streams 120 are rare, e.g. [11.23], [11.24].

When comparing these results with those for invertebrates, the latter were present in 369 publications (from 1978 until the end of 2022), with this discrepancy revealing the long way to go in diatom research in these ecosystems. Also [11.3] reviewed the literature on Intermittent Rivers (IR), highlighting the increasing characterization of these systems, considering theoretical aspects, assessment, protection, and habitat restoration. Nevertheless, in their review, only invertebrates and fish were considered among the biota of IR, along with biogeochemistry and hydrological and ecological assessment.

128 [Figure 11.3 here]

Figure 11.3. Total number of publications on non-perennial watercourses (light grey)and including diatoms (dark grey)

The few publications on algae, and particularly diatoms, do not reflect their essential role 131 in the functioning of temporary rivers, as the basis of the trophic network along with the 132 other primary producers (e.g. cyanobacteria, mosses, and macrophytes in the river 133 134 channel and the riparian vegetation in the margins). These organisms produce organic matter using upstream and lateral inorganic nutrients and carbon dioxide in the 135 136 photosynthesis process with solar energy [11.25]. In riverine systems, primary producers 137 differ, with the riparian vegetation dominating the terrestrial-aquatic ecotone, whilst the 138 aquatic environment is colonized by macrophytes and algae [11.25].

The structure of algal assemblages is commonly determined by the type of substratum, the water chemistry, light availability, water temperature, and flow [11.26]. In temporary rivers, these environmental factors are coupled with the challenges associated with the hydrological fluctuations, including the long dry periods and the hydric and thermal stress [11.25].

In this chapter we review 1) the biological aspects of diatoms in temporary rivers, focusing on their adaptation mechanisms to the fluctuating hydrology and associated stressors, such as flow reduction/lentification, increased temperature, desiccation, and UV radiation exposure; 2) diatom community dynamics and the importance of temporary rivers for endangered species conservation; and 3) diatom as bioindicators in temporary rivers.

150

151 **11.2** Diatoms adaptation mechanisms in temporary rivers

Temporary rivers represent a harsh and rigorous environment for diatoms, due to the hydric stress, related to drying and thermal stress, water flow fluctuations, atmospheric exposure, and desiccation. Globally, water stress directly affects cell structure, biomass, and algal diversity, since water is essential for phototrophs because it is involved in the cells' osmotic equilibrium, in photosynthesis as a reducing agent and in sugar synthesis, and in other metabolic processes [11.25]. Further, vegetative cells of benthic freshwater diatoms are known to be highly sensitive to desiccation, freezing, and abrupt heating, whilst vegetative cells of terrestrial diatoms are more tolerant to temperature extremes[11.27].

Diatoms have developed adaptation mechanisms (either life cycle, morphological or 161 physiological traits) related to the capacity to withstand these harsh conditions and persist 162 163 in the stream (by staying in, or colonizing potential refuges as pools, leaf litter packs, or subsuperficial sediments) and recover and spread once the flow resumes [11.9]. However, 164 165 short evidence of diatom adaptation mechanisms is yet available from temporary rivers, 166 as seen in Table 11.1. Thus, in this chapter information about the possible adaptation 167 mechanisms to the stressors diatoms face in temporary rivers is compiled but includes 168 data available from other ecosystems.

169 [Table 11.1 here]

Morphological and physiological changes in freshwater diatoms exposed to adverse conditions (e.g., temperature, light, desiccation, nutrients, pH, or salinity) have been reported [11.28], namely, the formation of resting stages, defined by [11.29] as cells or group of cells that stop growing and have their metabolic processes at a minimum. Two types have been described [11.27]:

- (i) resting spores (hypnospores), that differ in morphology and physiology from
 vegetative cells, with thicker frustules, often a rounder shape, and less
 elaborated patterns at the cell surface [11.28];
- 178 (ii) resting cells, with cytoplasmatic and physiological alterations but with similar
 179 morphology to the vegetative cells, a fact that can contribute to their
 180 overlooking.

181 Spores occurrence has been more common in marine centric diatoms, with few freshwater species as exceptions, mainly belonging to the genera Urosolenia and Acanthoceras 182 183 [11.30], Aulacoseira and inland Chaetoceros [11.28], and being rare amongst pennate diatoms, with few exceptions, such as Craticula cuspidata (Kützing) Mann, whose 184 185 resting-spore formation is detailed in [11.31]. Resting cells are more often formed in freshwater and pennate diatoms [11.28] and their existence explains the persistence of 186 187 viable cells where spores are not found (for instance, a sample collected in Southern Portugal from cobbles that were dry for a long time, at the end of the summer, revealed a 188 189 high proportion of live diatoms after rewetting in the laboratory). In diatoms, resting 190 stages are formed vegetatively e.g. [11.32], with a few exceptions of sexual spore 191 formation inside the auxospores in a few marine diatoms, e.g. *Leptocylindrus danicus*192 Cleve, *L. minimus* Gran [11.28].

193 Resting stages are reported to withstand unfavourable conditions for years, decades, and 194 even millennia and still be alive, e.g. (i) Aulacoseira granulata (Ehrenberg) Simonsen 195 resting cells present in anoxic sediments for twenty years were still able to germinate [11.33]; (ii) ~6600 cal. year BP old Chaetoceros muelleri var. subsalsum (Lemmermann) 196 197 Johansen & Rushforth resting spores in hypoxic sediments were still viable (even though the germination rate decreased with increasing age, from three hours to 2 - 3 days) 198 199 [11.34]. According to [11.28], cell survival in the resting stage depends on several 200 important factors, as light, temperature, and time.

[11.28] listed 16 non-marine taxa as producing resting spores and 17 producing restingcells:

(i) resting spores producing taxa - Acanthoceras zachariasii (Brun) Simonsen, 203 Aulacoseira bellicosa (Héribaud) Simonsen, A. italica (Ehr.) Simonsen, A. 204 skvortzowii Edlund, Stoermer et Taylor, Craticula cuspidata (Kützing) Mann, 205 206 Diatoma anceps (Ehrenberg) Kirchner, Eunotia faba (Ehrenberg) Grunow, E. pectinalis (Kützing) Rabenhorst, E. soleirolii (Kützing) Rabenhorst, Melosira 207 208 charcotii Peragallo, M. laevis (Ehrenberg) Ralfs, M. semilaevis Grunow, M. 209 turgida Ehrlich, Meridion circulare (Greville) C.Agardh, Urosolenia eriensis 210 (H.L. Smith) Round et Crawford, and U. longiseta (Zacharias) Edlund & 211 Stoermer;

(ii) resting cells producing taxa - Actinocyclus normanii (Gregory ex Greville) 212 Hustedt, Asterionella formosa Hassall, Aulacoseira granulata (Ehrenberg) 213 Simonsen, A. islandica (O.Müller) Simonsen, A. subarctica (O. Müller) 214 11. Haworth, Diatoma tenuis C.Agardh, Fragilaria capucina Desmazières, 215 Staurosira construens Ehrenberg, Fragilaria crotonensis Kitton, F. 216 217 intermedia Grunow, F. pinnata Ehrenberg, Stephanodiscus alpinus Hustedt, S. binderanus (Kützing) Krieger, S. medius Håkansson, S. niagarae 218 219 Ehrenberg, Tabellaria fenestrata (Lyngbye) Kützing, and T. flocculosa (Roth) 220 Kützing.

Physiological distinctions between resting stages and vegetative cells have been observed
and are reported in [11.28], namely: (i) the higher rates of cellular carbon to nitrogen; (ii)

higher carbon and chlorophyll content per cell when in the dark; (iii) more condensed 223 organelles; (iv) more or larger vesicles of storage products; (v) lower sugar phosphate 224 content; (vi) decrease in the respiration rate (to less than 20% when compared with 225 actively growing cells); (vii) reduction in the photosynthesis rate (less than 4% to that in 226 227 vegetative cells), related to the chloroplasts size reduction, loss, or colour change (from green to yellow and orange-red), observed in diatom cells during the non-flow phase by 228 229 [11.26]. An accumulation of oil in the cells of *Pinnularia viridis* was also observed by [11.29] in experimental drying. According to [11.29], Pinnularia viridis (Nitzsch) 230 Ehrenberg, P. major (Kützing) Rabenhorst, Cymbella lanceolata C. Agardh, and 231 Nitzschia linearis W. Smith were species mainly found in the water, but able to survive 232 some exposure and drying, while Nitzschia palea (Kützing) W.Smith and Stauroneis 233 234 phoenicenteron (Nitzsch) Ehrenberg were able to survive prolonged and severe drought.

Table 11.1 summarizes the adaptation mechanisms to the stressors in temporary rivers.

236

237 11.3 Flow reduction / lentification

In temporary rivers, one of the environmental factors most challenging for biological communities is flow reduction, which can be the flow interruption or even the complete drying of the channel, according to the Aquatic Regime (AR) of the temporary rivers, e.g. temporary-pools, temporary-drying or episodic, following the updated classification proposed by [11.35], that describes the long-term global variability of the hydrological conditions.

In temporary rivers, the duration and predictability of flow and no-flow periods are major
factors shaping biological communities [11.35].

Considering the biological communities' dependence on the changes of the aquatic habitats over time, [11.35] developed a methodology to analyse the aquatic regimes based on Aquatic States (AS), that relate to the sets of mesohabitats occurring in a reach at a determined moment, depending on the hydrological conditions (presented in Figure 11.4):

i) Hyperrheic – high water (flood), that originates the movement of stream bed
alluvium and drift of most of the aquatic fauna;

- 252 ii) Eurheic optimal hydraulic connectivity and widest range of discharges due
 253 to the succession of riffles and pools, presence and connectivity of all
 254 mesohabitats;
- 255 iii) Oligorheic low water discharge, but enough to connect most pools due to
 256 water rivulets;
- iv) Arheic surface discharge close to 0, but some disconnected pools are still
 present in the stream bed;
- v) Hyporheic absence of surface water in most of the stream bed, but the
 alluvium may be still close to saturation;
- vi) Edaphic no surface water in the entire stream bed, and the dryness of the
 alluvium does not allow active hyporheic life.

The importance of these aquatic states for the ecological status assessment is also discussed by [11.35], based on macroinvertebrate communities.

265 [Figure 11.4 here]

Figure 11.4. Aquatic States (AS), depending on the hydrological conditions (based on
[11.35])

Further, the hypothesis that also diatom communities respond to the AS was validated by 268 269 [11.36], based on a one-year study of permanent (as control) and temporary rivers in 2017 in Southern Portugal. 2017 was a very dry year, characterized by severe droughts, with 270 271 low accumulated precipitation even in winter and resuming only in March 2018. Therefore, the arheic state was distinguished between summer and winter, with 272 273 temperature being the main environmental factor separating the two types. Diatom 274 communities not only differed between AS (arheic and eurheic), but also reflected the 275 mesohabitats (flowing water and pools). Indicator species of the AS (eurheic in spring 2017, eurheic during the remaining seasons, arheic in summer, arheic in the remaining 276 seasons) were also retrieved and are as follows: 277

- arheic (except Summer 2017) - *Craticula cuspidata* (Kützing) D.G.Mann, *Fallacia pygmaea* (Kützing) Stickle & D.G. Mann, *Navicula erifuga* Lange-Bertalot, *Nitzschia intermedia* Hantzsch ex Cleve & Grunow, *Nitzschia supralitorea* Lange-Bertalot, *Planothidium pericavum* (J.R.Carter) Lange-Bertalot, *Sellaphora pupula* (Kützing)
Mereschkowksy, and *Surirella angusta* Kützing;

- arheic in Summer 2017 - Achnanthidium eutrophilum (Lange-Bertalot) Lange-Bertalot, *Cocconeis placentula* Ehrenberg, *Eolimna minima* (Grunow) Lange-Bertalot, *Epithemia turgida* (Ehrenberg) Kützing, *Karayevia clevei* (Grunow) Bukhtiyarova, *Nitzschia amphibia* Grunow, *N. filiformis* var. *conferta* (Richter) Lange-Bertalot, *N. valdestriata*Aleem & Hustedt, *Pseudofallacia tenera* (Hustedt) Liu, Kociolek & Wang, *Pseudostaurosira brevistriata* (Grunow) D.M.Williams & Round, *Pseudostaurosiropsis*sp. 1, and *Staurosirella pinnata* (Ehrenberg) D.M.Williams & Round;

- eurheic - Denticula subtilis Grunow, Encyonema minutiforme Krammer, Eunotia *implicata* Nörpel, Lange-Bertalot & Alles, E. minor (Kützing) Grunow, E. pectinalis
(Kützing) Rabenhorst, Fragilaria aff. capitellata (Grunow) J.B.Petersen, and Nitzschia
gracilis Hantzsch;

- eurheic in Spring 2017 - two Fragilaria species with affinity to the pectinalis group, *Gomphonema pumilum* var. *rigidum* E.Reichardt & Lange-Bertalot, *G. truncatum*Ehrenberg, *Melosira varians* C.Agardh, *Planothidium* cf. *frequentissimum* (LangeBertalot) Lange-Bertalot, and *Ulnaria acus* (Kützing) Aboal.

These assemblage shifts were also reported by [11.25], who stated that algal assemblages during the flowing phase were similar to permanent streams. With intermittency, a shift in the communities is observed, with lotic (benthic) algae gradually being replaced by lentic communities, with a dominance of the motile form in the pools, related to the nutrient increase and siltation [11.36]. When complete drying does not occur, these pools play an important role in the recolonization process once the flow resumes [11.37].

304

305 11.4 Desiccation stress

Water supports vital biophysical and biochemical processes and influences key cellular 306 307 functions, such as: protein folding and stability, enzyme catalysis, DNA packaging, molecular recognition, cellular tolerance to freezing, intracellular signalling and transport 308 309 processes; also biomolecules such as nucleic acids, proteins or membranes are normally 310 in close contact with water solutions [11.38]. Further, alterations in cell organelles 311 morphology and location have been observed, as water leaves the cell during the desiccation process, e.g. the shrinking of the protoplasmic components and distortion of 312 313 chloroplasts and pyrenoids of Stauroneis anceps cells, with dry cells organelles

positioned at the ends of the frustule, with the entire protoplast at one end or divided andoccupying both ends of the frustule [11.39].

316 During the dry phase, biomolecules face air drying/dehydration, which is the process involving the removal of both freezable and non-freezable fractions of water, with 317 318 extreme dehydration often causing: protein denaturation (a process that may be 319 irreversible); destabilization or disruption of membranes [11.40]; and the accumulation 320 of reactive oxygen species [11.41]. Tolerance to desiccation has been mainly observed in 321 resting cells of terrestrial diatoms by [11.27], with some mechanisms already identified 322 at the cellular level that may contribute to achieving it: (i) the addition of a solute 323 (disaccharides) can contribute to proteins' stabilization, and therefore dehydration to 324 much lower water content. In brief, as dehydration progresses, the solute concentration 325 increases to the point where it penetrates the protein hydration shell (one or two layers of 326 water molecules that are in close interaction with the surface of the proteins [11.38], 327 allowing the preservation of the conformational state of the dry protein [11.40]; (ii) further, sugars (namely the disaccharide trehalose) are effective in the stabilization of 328 329 phospholipid bilayers under severe dehydration, allowing dry phospholipids to be in liquid crystalline phase at temperatures they would normally be in the gel phase, avoiding 330 the gel to the liquid crystalline phase transition during rehydration [11.40], [11.42]. 331 Further, according to the principles of the excluded solute theory, all compatible solutes 332 (low molecular mass organic solutes) would be expected to provide some protection to 333 cells and intracellular macromolecules, against the reduced intracellular water potential 334 335 and the resulting increase in intracellular ionic strength caused by the water loss [11.42]. 336 However, under a severe desiccation state, this mechanism is not efficient, since ca. 0.3 -0.4 g H₂O protein⁻¹ are needed to maintain even a single mono-layer of bound water 337 around the proteins [11.42]. 338

339 Diatom response in temporary rivers to drying includes the prevalence of the life-forms mobile and mucilage-forming [11.25]. The first trait includes species that are able to move 340 341 ("mobile") and are therefore capable of selecting the most suitable habitat [11.43] and migrating, for instance, to deeper sediments where moisture can remain [11.9], [11.44], 342 343 or to exploit refugia (as pools). Mucilage-forming species (e.g. Cymbella or Gomphonema species) form mucilaginous stalks or tubes that can protect the cells. The 344 345 increase in the production of extracellular substances in stress situations is known to occur 346 when there is an inhibition of cell division but photoassimilation continues [11.45].

These adaptations, either as resting stages or certain life-forms prevalence, allow diatom 347 communities to persist in dry streambeds only when the drying process is recurrent and 348 slow [11.25], resulting from a long evolutionary period during which the multihabitat 349 filter process selects diatoms with a set of traits that allows their adaptation [11.46], as in 350 351 Mediterranean temporary rivers. This acclimation process has been experimentally tested by [11.27], [11.39]. The first authors acknowledging that *Stauroneis anceps* survival is 352 353 increased by slowing drying; and the latter proving that gradual heating before desiccation had positive results only on the survival of resting cells of terrestrial diatoms, that have 354 355 protection mechanisms to survive in soils and temporary water bodies, being regularly 356 exposed to harsh changes in humidity and temperature.

When diatom communities are subjected to rapid alterations in natural flow conditions, i.e. magnitude, frequency, duration, timing, and predictability [11.46], community recovery depends on their capacity to remain in drought refuges (e.g., permanent pools, moss patches, leaf packs, seeps and woody debris, dry biofilm on stones, dry sediments and leaf litter), from where regrow or recolonization may occur [11.37].

Adaptations to desiccation play a major role in shaping diatom communities in view of 362 the increasing magnitude, frequency, occurrence and duration of river drying, driven by 363 climate change and water abstraction [11.5], [11.12], [11.14], [11.47]. An increase in the 364 365 length of dry rivers with consequent connectivity loss has already been observed in 366 diverse regions due to the streamflow decrease and drought caused by a decrease in 367 precipitation and temperature increase [11.6]. Further, predicted temperature increase will 368 lead to an increase in evapotranspiration pushing systems closer or beyond the balance where water losses to the atmosphere exceed inputs. The probability of river drying is 369 370 also expected to increase due to the higher probability of dry periods occurrence, i.e., seasonal or multi-year droughts [11.5]. 371

372

373 11.5 Thermal stress

In temporary rivers, temperature increase occurs during the lentification process, as observed in Southern Portugal, where a maximum of 33.1 °C was measured in a pool at the end of the Summer (Ameira stream, 22nd September 2017), with the median for all the summer pools (11 sites) being 22.4 °C (Interquartile range: 18.35-28.15 °C), whilst the flowing streams (2 sites) presented lower temperatures (median: 14.60 °C; interquartile range: 13.95-15.25 °C) as depicted in [11.36]. Thus, diatoms are also
exposed to thermal stress during this dry phase, an important issue considering that
temperature is a relevant environmental factor that regulates algal growth and species
distribution, either seasonally, altitudinally or latitudinally [11.48]–[11.50].

Diatom specific growth in response to temperature depends on both the ranges and temperature-specific dependency [11.50], which is more evident at low (< 10 °C) and high temperatures (> 20 °C), but can be modified by light flux density and photoperiod [11.48].

387 Species growth rate is known to increase up to a temperature maximum, followed by an abrupt decrease until reaching the upper limit of the growth range, therefore, the 388 389 maximum rate occurs near the upper temperature range [11.50]. Hence, according to the 390 optimum conditions (with a higher relative growth rate) tendentially occur at the temperature each species predominates in the field [11.50]. In the absence of data from 391 temporary rivers, [11.50] study with planktonic diatoms in tem11.perate climate is 392 393 presented as an example. In this case, temperature dependency was higher in hightemperature species and lethal heating was 5 °C above the higher limit of the growth range 394 of the species, with the highest value of 40 °C for Chaetoceros pseudocurvisetus Mang., 395 Stephanodiscus hantzschii Grun. and Skeletonema costatum (Grev.) Cleve, allowing most 396 of the species to grow under the annual habitat temperature range. Several freshwater 397 planktonic algae, including diatoms, were not able to withstand temperatures above 30 °C 398 399 even though they were actively growing at 25°C (e.g., Asterionella formosa Hassall, 400 Tabellaria flocculosa var. asterionelloides (Grunow) Knudson, Aulacoseira subarctica (O.Müller) E.Y.Haworth and Discostella pseudostelligera (Hustedt) Houk & Klee), 401 402 except for Fragilaria crotonensis Kitton that was still actively growing at this temperature, but for which 35°C were lethal [11.48]. Furthermore, in the experiments 403 carried out by [11.51], abrupt heating up to 40 °C was lethal for 40% of the 34 studied 404 species, whilst more species were able to withstand gradual heating. 405

The stress response to high temperatures is considered to be a universal and highly conservative mechanism, that is typically characterized by a decrease in the production of constitutive proteins coupled with an increase in stress-inducible proteins (heat shock proteins, HSP). This response can be observed, for instance in the diatoms *Chaetoceros muelleri* [11.52], *Nitzschia alba* [11.53], and *Skeletonema costatum* [11.54], with a faster response to temperature increase detected in thermo-tolerant species (e.g. *Chaetoceros muelleri*) possessing higher amounts of constitutively expressed HSPs.

Additionally, changes in diatom fatty acid composition have been reported in response to
temperature increases above the optimal growth temperature level, triggered by a
particular temperature and not by the stress duration [11.55].

416 The exposure of Skeletonema costatum to heat stress (25°C – 40 min.) has caused a rapid 417 increase in free polyamine content (putrescine and spermidine titers), which can be explained by their release from the bound form (as a "first aid" against the initial shock), 418 419 followed by a new synthesis of putrescine after 30°C – 40 min [11.54]. The polycationic nature of polyamines seems to provide the cells with the ability to regulate the 420 421 intracellular environment, to have a protective effect on membrane integrity by putrescine 422 binding to phospholipids or the stabilization of other cell molecules by inhibition of degradative enzymes [11.54]. Also, conjugated polyamines are hypothesized to be 423 involved in the cellular response to heat stress (e.g. TCA-soluble bound spermine in 424 425 Skeletonema costatum) [11.54].

Thermotolerance of enzymes can also be enhanced by the presence of compatible solutes, since they are preferentially excluded from the immediate hydration shell of proteins, consequently, their presence in high concentrations would tend to inhibit the thermal denaturation of proteins [11.42].

Diatom's dependency on temperature and sensitivity to abrupt temperature changes make 430 them a highly sensitive group to climate change, with the predicted temperature increase, 431 432 alteration in precipitation patterns, and increase in the frequency and duration of extreme events such as droughts and floods. These extreme climate events can compromise the 433 434 conditions beyond which many species are adapted [11.21] [11.56]. Climate change impacts on freshwater ecosystems have already been detected at a global scale, including 435 shifts in species range and in timing. Also, at the 1.5°C global warming scenario, species 436 and ecosystems (mainly in polar and already-warm areas, where most of the non-437 438 perennial watercourses occur), will face temperatures beyond their historical values [11.56], that can severely alter freshwater diatom distribution, favouring species already 439 adapted to more harsh habitats (e.g. terrestrial diatoms) and even impairing their survival, 440 with consequences for non-perennial watercourses functioning. 441

442

443 11.6 UV Radiation exposure

Solar radiation is the electromagnetic radiation emitted by the sun and comprises a wide range of wavelengths, known as the electromagnetic spectrum, from radiowaves through the infrared, visible and ultraviolet to X-rays and gamma rays. However, 99% of the energy of solar radiation is comprised in the wavelength band from 150 to 4 000 nm (near ultraviolet, visible, and near-infrared regions of the solar spectrum).

According to the UCAR COMET Program, ca. 43% of radiant energy from the sun is in the visible part of the spectrum (between 400 – 700 nm), roughly 49% is infrared (between 700 nm -1 mm), about 7% is from ultra-violet (between 100 – 400 nm), and less than 1% of solar radiation is emitted as x-rays, gamma rays and radio waves (https://sos.noaa.gov/catalog/datasets/climatebits-solar-radiation/).

454 Considering that shorter wavelength radiation is more energetic, ultraviolet is the range that rises more concern, due to the possible damage it may cause in living organisms. It 455 is separated into three ranges: UV-C (100-280 nm), which makes up only 0.5% of all 456 solar radiation, and very little reaches the surface because it is absorbed by ozone; UV-B 457 (280-320 nm) is an energetic, photoactivating band of radiation, only partially absorbed 458 in the stratosphere, that can penetrate the water reaching 20 m depth in the ocean; UV-A 459 (320-400 nm) is not absorbed by ozone, though it can be blocked by cloud cover and can 460 penetrate deeper in the water than UV-B (Fondriest Environmental, 2014, 461 https://www.fondriest.com/environmental-measurements/parameters/weather/solar-radiation/). 462

Under climate change scenarios, drying increase in temporary rivers, both in length,
extension, frequency, and duration, may affect the exposure of diatoms to radiation,
including photosynthetically active radiation (PAR) and ultraviolet (UV). Thus,
organisms in these exposed areas should possess efficient adaptation mechanisms [11.51].

467 UV-B irradiance can be hazardous to living beings, as it affects biomolecules directly or by inducing the formation of reactive oxygen species (ROS, as hydrogen peroxide and 468 superoxide radicals) inside the cell. ROS can further affect the cellular structures, block 469 470 enzymatic reactions, interfere with photosynthesis (through changes to the main photosynthetic enzyme, ribulose bis-phosphate carboxylase), the nutrient uptake, DNA 471 replication and physiological responses as motility and orientation [11.57]–[11.61]. Thus, 472 at the cellular level, UV radiation toxic effect can be triggered in two photochemical 473 pathways: (i) direct or primary mechanisms - biomolecules as proteins and nucleic acids 474

have chromophores that absorb in the UV region of the radiation spectrum and are 475 photochemically degraded or transformed under high UV flux, impairing or losing their 476 biological function; (ii) indirect mechanisms - intermediate compounds absorb UV 477 478 (photosentising agents) either inside or outside the cell, and produce reactive oxygen 479 species (ROS). These high energy oxidants as hydrogen peroxide, superoxide or hydroxyl radicals can further react with other cellular components [57]. DNA and RNA are 480 especially sensitive to UV radiation because of their highest absorbance coefficients for 481 482 short wavelength UV [11.58], making genetic one of the most reported damages, also for 483 diatoms [11.57].

In the absence of specific information about diatoms in temporary rivers, the results of 484 485 [11.57] using cultured marine diatom species in laboratory experiments are addressed. Marine diatom species (Cyclotella sp., Nitzschia closterium and Thalassiosira 486 nordenskioldii) exposed to an increasing range of daily doses of UV-B (280-320 nm) 487 radiation revealed that, at the lowest dose (< 2000 J m⁻² d⁻¹, DNA weighted biologically 488 effective dose, normalized at 300 nm: daily BED_{DNA 300 nm}), division rates decreased, 489 volume enlargement occurred and high cellular protein and pigment content levels were 490 491 detected, including the photoprotective carotenoids (diadinoxanthin + diatoxanthin), or both, in the Cyclotella species. The highest UV-B treatments (between 2000 and 3800 J 492 m⁻² d⁻¹ daily BED_{DNA 300 nm}) resulted in complete growth inhibition, with minor changes 493 494 in protein, pigments and cell volume [11.57]. Further, high UV-B radiation levels induced 495 plasmolysis and disorientation of cell organelles in Cyclotella sp., whilst lower levels 496 seemed to increase chloroplasts in number and volume [11.57]. Thus, these results support the hypothesis that UV-B radiation causes growth without cell division, due to 497 498 DNA damage and the arrest in the S or G2 phase of the cell cycle [11.57]. Most microalgae developed strategies to minimize the UV-induced damage. According to 499 500 [11.62], these include:

501 502

503

 (i) avoidance mechanisms, by controlling their movements towards light or water surface, as is the case of motile diatoms, that can migrate to less exposed areas [11.61];

(ii) screening - reduction of the effective UV radiation that penetrates the cell, by
synthesising compounds that absorb the damaging wavelengths (screening
agents), either intracellular, as the mycosporine-like amino acids (MAAs),
also reported by [11.63] as being involved in the defence strategy in response

508to UV-induced damage in the diatom *Porosira glacialis*) or extracellular (at509the cell walls where they block incoming UV radiation);

- repair of UV-induced damages to the nuclear material (the most important 510 (iii) damages, for which cells have developed repair mechanisms generally 511 512 involving biosynthetic pathways mediated by stress proteins), proteins and enzymes, pigments and photosynthetic reaction centres, lipids and 513 membranes. In the case of DNA repair, several mechanisms have been 514 identified as photoreactivation (light-induced repair), nucleotide and base 515 516 excision repair and recombinational repair; and the activation of heat shock proteins for protein damage repair. For the UV damage caused by ROS, 517 518 detoxifying mechanisms have been developed, by means of antioxidant enzymes, lipid-soluble antioxidants in cellular membranes (e.g. carotenoids as 519 diadinoxanthin and beta-carotene), and water-soluble reductants in the 520 cytosol; 521
- 522 (iv) acclimation - the process by organisms adapt to changes in the environment, either by physiological changes or by alterations in the species composition at 523 524 the community level. Recent research also verified the hypothesis that the 525 optical properties of centric diatoms frustule can protect the cell from harmful radiation (UVB and UVC), and three main mechanisms involved in cell 526 protection were identified, using Coscinodiscus wailesii and C. cf. radiatus as 527 528 a model [11.61], [11.64]. These are: absorption by silica and trace organic compounds incorporated in the frustule; diffraction (due to the geometry of 529 the valve and refractive index contrast with respect to the surrounding 530 environment); and photoluminescence (mainly due to chemical surface 531 defects of hydrated amorphous porous silica). This hypothesis arose from the 532 533 fact that diatoms evolved under low concentrations of atmospheric oxygen and ozone and are characterized by high tolerance to UV irradiation, even with 534 535 low concentrations of screening agents such as MAAs [11.61], [11.64]. 536 Nonetheless, it could be interesting to verify it in pennate diatoms, since the test with Navicula perminuta was not conclusive due to its small dimensions 537 and high curvature [11.64]. Thus, this distribution of UV radiation due to their 538 SiO₂ frustules can be an important evolutionary aspect to be considered for 539 the presence and evolution of frustules in diatoms, contributing to a decrease 540 541 in the rate of UV radiation-induced damage in the cells [11.64]. Further, in the

review made by [11.60] it is concluded that research into the role of diatom
frustules in UV protection shows great potential, both in relation to diatom
biology and potential technological applications, in spite of the few number
of diatom species analysed so far.

546 The development of these strategies, is related with the previous light history (e.g. for the 547 development of UV screening) and cell size, for instance in the production of MAAs 548 [11.62], and a differential response to UV radiation was observed for two diatoms from 549 contrasting niches, planktic (collected offshore) and benthic (collected from intertidal 550 sediments. Benthic species presented higher tolerance to extreme temperature and UV 551 radiation) and a better response was observed under elevated temperature [11.65]. Given 552 these environmental-specific responses, it would be interesting to carry out detailed 553 studies on the adaptation mechanisms to UV exposure from diatoms in temporary rivers.

554 Thus, as an overview, UV exposure net stress reflects the balance between damage, repair and the energetic costs of protection, and can be presented in terms of an increase in 555 556 energy demand, changes in cell composition, and a decrease in growth and survival rates, as depicted in the scheme in [11.58]. Given that the physiological changes observed in 557 response to the increase in UV-B radiation do not confer full protection to the organisms, 558 and a number of organisms have already reached their UV-B tolerance limits [11.62], the 559 560 diatom frustule UV protection detailed in [60] may confer them an additional advantage in temporary rivers, more susceptible to drying. 561

562

563 11.7 Communities dynamics in temporary rivers

564 Community response to perturbations depends on the environment where it is developed, 565 with low-stress environments supporting complex communities, but fragile, when 566 compared to those subjected to environmental stress, e.g. pollutants exposure, light, 567 current or desiccation, as reported in the diverse studies on diatom communities cited by 568 [11.45].

According to [11.25], flow intermittence is a selective force on algal community composition, usually causing a decrease in alpha diversity. Indeed, a long drying phase can eliminate most of the sensitive species, few species are able to resist drying and there are few pioneer species able to recolonize when flow resumes. Thus, flow intermittency also alters diatom composition throughout the year, with shifts from lotic to species adapted to lentic conditions, even up to aerophilous species during the lentification process and drying up of the riverbed [11.66], [11.67]. Nevertheless, when drying is not complete, beta diversity is higher than expected, since lotic, lentic, and terrestrial habitats cooccur sequentially in a longitudinal direction [11.25].

578 Further, according to [11.68], diatom communities in temporary rivers have less species 579 and a lower proportion of specialist taxa than perennial streams. Also [11.66] detected 580 taxonomic and functional differences between temporary and permanent reaches in 581 Liguria (NW-Italy), both at local and regional scales, with the temporary reaches being 582 characterized by less heterogeneous communities, with higher percentages of small, 583 mainly stalked and pioneer taxa belonging to the low profile guild, which are able to 584 tolerate physical disturbance. These results are in accordance with [11.69], in their study 585 of intermittent and permanent lowland rivers in Carpathian basin. The authors observed 586 that intermittent rivers were characterized by lower taxonomical diversity, higher Berger-587 Parker index (a metric to measure dominance, defined by the proportional abundance of 588 the dominant species in the assemblage), and lower functional richness and diversity. Disturbed sections were dominated by small-sized generalists (including pioneers and 589 590 small low-profile taxa) and aerophilic taxa, the latter considered good indicators of 591 drought, since they are able to survive the drying up of the riverbed and colonise rapidly 592 the habitats once flow resumes. The importance of hydrology-related variables in 593 determining diatom species distribution and community ecological attributes in Cyprus was also acknowledged by [11.71]. Further, even in large rivers, a continuous decrease in 594 precipitation (drying period) significantly decreased benthic diatoms, in the number of 595 596 taxa, Shannon diversity, and functional richness [11.70], addressing the implications of 597 the longer periods with low precipitation, as predicted in the climate change scenarios.

598 Temporary rivers' importance for species conservation is another issue that should be 599 highlighted, thus reinforcing the need for the adoption of appropriate conservation and 600 management measures. This is supported by the relevance and frequency of species that 601 fulfil the descriptions of Red List threat categories and that are presented in the diatom 602 Red List for Central Europe (2018) in intermittent streams in Cyprus [11.71]. This is in 603 accordance with the findings of [11.36] and [11.72], who highlighted the fact that flow instability, lentification and habitat fragmentation represented the major threats for 604 605 endangered species, whilst water stability and river habitat heterogeneity favour their 606 presence. These results highlighted the importance of the isolated pools as refugia for benthic diatoms during the dry phase, especially under the climate change scenarios, with
increasing drought causing the persistence of isolated pools beyond the summer season
(e.g. in 2007 Southern Portugal was subjected to a severe drought, with isolated pools
permanence during the winter until flow resuming only in the next spring).

611

612 11.8 Diatoms as bioindicators in temporary rivers

613 Diatoms have been largely used as bioindicators in the framework of the monitoring in accordance with the WFD [11.17], being commonly used as proxies for the Biological 614 615 Quality Element - Phytobenthos [11.73]. However, for the ecological status assessment 616 diatom sampling should be carried out in stable flow conditions [11.74], avoiding zones 617 of very slow current [11.75]. The fact that the dry phase is excluded gives an incomplete idea of the habitat quality in temporary rivers [11.76], thus, complementary methods for 618 the ecological status assessment should be designed and tested. Further, an adaptation or 619 620 development of novel metrics to assess the ecological status in temporary rivers is envisaged, since metrics developed for permanent streams may underestimate their 621 ecological status since sensitivity to intermittency and to environmental degradation often 622 623 covary [11.76]. Diatoms have the potential to fill this lacune, as presented in the 624 preliminary results depicted in [11.76] and further tested by [11.36], with the Specific pollution sensitive index (SPI) negatively correlated with the percentage of dry riverbed 625 626 and presenting a good response over time, independently of the aquatic state The Ecological Quality Ratio (EQR) for dry biofilm samples collected in Summer was 627 628 comparable with the values from the previous spring, thus dry biofilm can be considered a viable alternative for the ecological status assessment during the dry phase, instead of 629 630 the isolated pools, whose communities are not considered by [11.35] to be representative 631 of the ecological status of the stream. Also, [11.77] observed that the Biological Diatom 632 Index (BDI) was slightly affected by drying, but these effects had no significant influence 633 on the ecological quality assessment. [11.78] also validated this hypothesis, in a study conducted in small hilly streams in the Pannonian ecoregion (Hungary), with drying only 634 affecting negatively the Rott's trophic index (TID), whilst the Specific pollution 635 sensitivity index (IPS), Rott's saprobic index (SID) and the Hungarian phytobenthos 636 637 metric (IPSITI) were not impacted.

638

639 11.9 Final remarks

Diatoms have a recognized importance in temporary rivers as the main algal component 640 of biofilms, with any changes in their taxonomical and functional diversity, physiology 641 or biomass being repercussed in the whole ecosystem. Furthermore, adaptation 642 643 mechanisms of diatoms have already been detected, and contribute to enhance their response to main environmental stressors (e.g. flow reduction, temperature increase, 644 increased exposure to UV radiation and desiccation), and reveal that diatoms can have an 645 adaptative advantage under the predicted climate change extreme conditions. However, 646 647 some of these strategies were tested with diatoms adapted to other aquatic ecosystems. Therefore, detailed studies should be carried out with diatoms adapted to naturally 648 intermittent rivers to validate their effectiveness to the particular conditions 649 650 characterizing these ecosystems. Further, diatoms can also play an important role in the 651 management of temporary rivers, since they can be used as bioindicators even during the dry phase. Appropriate management of naturally temporary rivers is a must since, apart 652 from their value and global importance, they also host fragile diatoms (still poorly 653 explored) and include threatened species. Further studies on diatoms in temporary rivers 654 655 should be addressed, since the ecology of most species inhabiting these ecosystems is still 656 poorly studied.

657

658 **References**

11.1 Magand, C., Helena Alves, M., Calleja, E., Datry, T., Dörflinger, G., & England,
J. (2020). *Intermittent Rivers and Ephemeral streams: What water managers need to know.* <u>https://doi.org/10.5281/zenodo.3888474</u>

11.2 Datry, T., Boulton, A. J., Bonada, N., Fritz, K., Leigh, C., Sauquet, E., Tockner,
K., Hugueny, B., & Dahm, C. N. (2018a). Flow intermittence and ecosystem services in
rivers of the Anthropocene. In *Journal of Applied Ecology* (Vol. 55, Issue 1, pp. 353–
364). Blackwell Publishing Ltd. <u>https://doi.org/10.1111/1365-2664.12941</u>

Leigh, C., Boulton, A. J., Courtwright, J. L., Fritz, K., May, C. L., Walker, R. H.,
& Datry, T. (2016). Ecological research and management of intermittent rivers: an
historical review and future directions. *Freshwater Biology*, *61*(8), 1181–1199.
https://doi.org/10.1111/fwb.12646

11.4 Tolonen, K. E., Picazo, F., Vilmi, A., Datry, T., Stubbington, R., Pařil, P., Perez
Rocha, M., & Heino, J. (2019). Parallels and contrasts between intermittently freezing
and drying streams: From individual adaptations to biodiversity variation. In *Freshwater Biology* (Vol. 64, Issue 10, pp. 1679–1691). Blackwell Publishing Ltd.
https://doi.org/10.1111/fwb.13373

Allen, D. C., Datry, T., Boersma, K. S., Bogan, M. T., Boulton, A. J., Bruno, D.,
Busch, M. H., Costigan, K. H., Dodds, W. K., Fritz, K. M., Godsey, S. E., Jones, J. B.,
Kaletova, T., Kampf, S. K., Mims, M. C., Neeson, T. M., Olden, J. D., Pastor, A. V., Poff,
N. L. R., & Zimmer, M. (2020). River ecosystem conceptual models and non-perennial
rivers: A critical review. In *Wiley Interdisciplinary Reviews: Water* (Vol. 7, Issue 5). John
Wiley and Sons Inc. https://doi.org/10.1002/wat2.1473

Allen, D. C., Kopp, D. A., Costigan, K. H., Datry, T., Hugueny, B., Turner, D. S.,
Bodner, G. S., & Flood, T. J. (2019). Citizen scientists document long-term streamflow
declines in intermittent rivers of the desert southwest, USA. *Freshwater Science*, *38*(2),
244–256. <u>https://doi.org/10.1086/701483</u>

11.7 von Schiller, D., Bernal, S., Dahm, C. N., & Martí, E. (2017). Nutrient and
Organic Matter Dynamics in Intermittent Rivers and Ephemeral Streams. In *Intermittent Rivers and Ephemeral Streams: Ecology and Management* (pp. 135–160). Elsevier Inc.
<u>https://doi.org/10.1016/B978-0-12-803835-2.00006-1</u>

- 689 11.8 Datry, T., Foulquier, A., Corti, R., Von Schiller, D., Tockner, K., Mendoza-Lera,
- 690 C., Clément, J. C., Gessner, M. O., Moleón, M., Stubbington, R., Gücker, B., Albarinõ,
- 691 R., Allen, D. C., Altermatt, F., Arce, M. I., Arnon, S., Banas, D., Banegas-Medina, A.,
- Beller, E., Zoppini, A. (2018b). A global analysis of terrestrial plant litter dynamics in
 non-perennial waterways. *Nature Geoscience*, *11*(7), 497–503.
 <u>https://doi.org/10.1038/s41561-018-0134-4</u>

695 11.9 Sabater, S., Timoner, X., Borrego, C., & Acuña, V. (2016). Stream biofilm responses to flow intermittency: From cells to ecosystems. In Frontiers in Environmental 696 697 Science (Vol. Frontiers Media S.A. 4, Issue MAR). 698 https://doi.org/10.3389/fenvs.2016.00014

11.10 Stubbington, R., England, J., Wood, P. J., & Sefton, C. E. M. (2017). Temporary
streams in temperate zones: recognizing, monitoring and restoring transitional aquaticterrestrial ecosystems. *Wiley Interdisciplinary Reviews: Water*, 4(4).
https://doi.org/10.1002/WAT2.1223

- 11.11 Raymond, P. A., Hartmann, J., Lauerwald, R., Sobek, S., McDonald, C., Hoover,
 M., Butman, D., Striegl, R., Mayorga, E., Humborg, C., Kortelainen, P., Dürr, H.,
 Meybeck, M., Ciais, P., & Guth, P. (2013). Global carbon dioxide emissions from inland
 waters. *Nature*, *503*(7476), 355–359. <u>https://doi.org/10.1038/nature12760</u>
- 11.12 Datry, T., Larned, S. T., & Tockner, K. (2014). Intermittent rivers: A challenge for freshwater ecology. In *BioScience* (Vol. 64, Issue 3, pp. 229–235). Oxford University Press. <u>https://doi.org/10.1093/biosci/bit027</u>
- 11.13 Messager, M. L., Lehner, B., Cockburn, C., Lamouroux, N., Pella, H., Snelder,
 T., Tockner, K., Trautmann, T., Watt, C., & Datry, T. (2021). Global prevalence of nonperennial rivers and streams. *Nature*, 594(7863), 391–397.
- 713 https://doi.org/10.1038/s41586-021-03565-5

11.14 Dhungel, S., Tarboton, D. G., Jin, J., & Hawkins, C. P. (2016). Potential Effects
of Climate Change on Ecologically Relevant Streamflow Regimes. *River Research and Applications*, *32*(9), 1827–1840. https://doi.org/10.1002/rra.3029

11.15 Acuña, V., Datry, T., Marshall, J., Barceló, D., Dahm, C. N., Ginebreda, A.,
McGregor, G., Sabater, S., Tockner, K., & Palmer, M. A. (2014). Why should we care
about temporary waterways? In *Science* (Vol. 343, Issue 6175, pp. 1080–1081). American
Association for the Advancement of Science. https://doi.org/10.1126/science.1246666

11.16 Stubbington, R., Acreman, M., Acuña, V., Boon, P. J., Boulton, A. J., England,
J., Gilvear, D., Sykes, T., & Wood, P. J. (2020). Ecosystem services of temporary streams
differ between wet and dry phases in regions with contrasting climates and economies.

- 724 *People and Nature*, 2(3), 660–677. <u>https://doi.org/10.1002/pan3.10113</u>
- 11.17 Directive 2000/60/EC. (2000). *I establishing a framework for Community action in the field of water policy*.
- 11.18 Steward, A. L., Negus, P., Marshall, J. C., Clifford, S. E., & Dent, C. (2018).
 Assessing the ecological health of rivers when they are dry. *Ecological Indicators*, *85*,
 537–547. <u>https://doi.org/10.1016/j.ecolind.2017.10.053</u>
- 11.19 Stubbington, R., Chadd, R., Cid, N., Csabai, Z., Miliša, M., Morais, M., Munné, 730 A., Pařil, P., Pešić, V., Tziortzis, I., Verdonschot, R. C. M., & Datry, T. (2018). 731 Biomonitoring of intermittent rivers and ephemeral streams in Europe: Current practice 732 and priorities to enhance ecological status assessments. In Science of the Total 733 Environment (Vol. 1096-1113). Elsevier B.V. 734 618, pp. https://doi.org/10.1016/j.scitotenv.2017.09.137 735
- 11.20 Acuña, V., Hunter, M., & Ruhí, A. (2017). Managing temporary streams and
 rivers as unique rather than second-class ecosystems. *Biological Conservation*, 211, 12–
 <u>https://doi.org/10.1016/j.biocon.2016.12.025</u>
- 11.21 Chiu, M. C., Leigh, C., Mazor, R., Cid, N., & Resh, V. (2017). Anthropogenic
 Threats to Intermittent Rivers and Ephemeral Streams. In *Intermittent Rivers and Ephemeral Streams: Ecology and Management* (pp. 433–454). Elsevier Inc.
 https://doi.org/10.1016/B978-0-12-803835-2.00017-6
- 11.22 Reid, A. J., Carlson, A. K., Creed, I. F., Eliason, E. J., Gell, P. A., Johnson, P. T.
 J., Kidd, K. A., MacCormack, T. J., Olden, J. D., Ormerod, S. J., Smol, J. P., Taylor, W.
 W., Tockner, K., Vermaire, J. C., Dudgeon, D., & Cooke, S. J. (2019). Emerging threats
 and persistent conservation challenges for freshwater biodiversity. *Biological Reviews*,
 94(3), 849–873. <u>https://doi.org/10.1111/brv.12480</u>
- 11.23 Stanish, L. F., O'Neill, S. P., Gonzalez, A., Legg, T. M., Knelman, J., Mcknight,
 D. M., Spaulding, S., & Nemergut, D. R. (2013). Bacteria and diatom co-occurrence
 patterns in microbial mats from polar desert streams. *Environmental Microbiology*, *15*(4),
- 751 1115–1131. <u>https://doi.org/10.1111/j.1462-2920.2012.02872.x</u>
- 11.24 Stanish, L. F., Kohler, T. J., Esposito, R. M. M., Simmons, B. L., Nielsen, U. N.,
 Wall, D. H., Nemergut, D. R., & McKnight, D. M. (2012). Extreme streams: Flow
 intermittency as a control on diatom communities in meltwater streams in the McMurdo
 Dry Valleys, Antarctica. *Canadian Journal of Fisheries and Aquatic Sciences*, 69(8),
 1405–1419. <u>https://doi.org/10.1139/F2012-022</u>

11.25 Sabater, S., Timoner, X., Bornette, G., De Wilde, M., Stromberg, J. C., & Stella,
J. C. (2017). The Biota of Intermittent Rivers and Ephemeral Streams: Algae and
Vascular Plants. In *Intermittent Rivers and Ephemeral Streams: Ecology and Management* (pp. 189–216). Elsevier Inc. <u>https://doi.org/10.1016/B978-0-12-803835-</u>
2.00016-4

11.26 Timoner, X., Buchaca, T., Acuña, V., & Sabater, S. (2014). Photosynthetic
pigment changes and adaptations in biofilms in response to flow intermittency. *Aquatic Sciences*, 76(4), 565–578. <u>https://doi.org/10.1007/s00027-014-0355-6</u>

11.27 Souffreau, C., Vanormelingen, P., Sabbe, K., & Vyverman, W. (2013). Tolerance
of resting cells of freshwater and terrestrial benthic diatoms to experimental desiccation
and freezing is habitat-dependent. *Phycologia*, 52(3), 246–255.
https://doi.org/10.2216/12-087.1

- 11.28 Mcquoid, M. R., & Hobson, L. A. (1996). Review diatom resting stages. In J.
 Phycol (Vol. 32).
- 11.29 Evans, J. H. (1958). The Survival of Freshwater Algae During Dry Periods: Part

I. An Investigation of the Algae of Five Small Ponds. In *Source: Journal of Ecology* (Vol.

773 46, Issue 1). <u>https://www.jstor.org/stable/2256910</u>

- 11.30 Edlund, M. B., & Stoermer, E. F. (1993). Resting spores of the freshwater diatoms
 Acanthoceras and Urosolenia. In *Journal of Paleolimnology* (Vol. 9).
- 11.31 Schmid A. (2009). Induction of resting-spores in the pennate diatom Navicula
 (Craticula) cuspidata by uncoupling of the cell and plastid cycles. Nova Hedwigia.
 [Online] 85–101.
- 11.32 Drebes, G. (1966). On the life history of the marine plankton diatom
 Stephanopyxis palmeriana. *Helgol. Wiss. Meeresunters.*, 13, 101–114.
- 11.33 Sicko-Goad, L., Stoermer, E. F., & Fahnensliel, G. (1986). Rejuvenation of
 Melosira granulata (Bacillariophyceae) resting cells from the anoxic sediments of
 Douglas Lake, Michigan. I. Light Microscopy and 14C uptake. In / *Phycol* (Vol. 22).
- 11.34 Sanyal, A., Larsson, J., van Wirdum, F., Andrén, T., Moros, M., Lönn, M., &
 Andrén, E. (2022). Not dead yet: Diatom resting spores can survive in nature for several
 millennia. *American Journal of Botany*, 109(1), 67–82. https://doi.org/10.1002/ajb2.1780
- 11.35 Gallart, F., Prat, N., Garca-Roger, E. M., Latron, J., Rieradevall, M., Llorens, P., 787 788 Barbera, G. G., Brito, D., De Girolamo, A. M., Lo Porto, A., Buffagni, A., Erba, S., Neves, R., Nikolaidis, N. P., Perrin, J. L., Querner, E. P., Quinonero, J. M., Tournoud, M. 789 G., Tzoraki, O., ... Froebrich, J. (2012). A novel approach to analysing the regimes of 790 791 temporary streams in relation to their controls on the composition and structure of aquatic 792 biota. Hydrology and System Sciences, 16(9), 3165-3182. Earth 793 https://doi.org/10.5194/hess-16-3165-2012
- 11.36 Novais, M. H., Morales, E. A., Penha, A. M., Potes, M., Bouchez, A., Barthès, A.,
- 795 Costa, M. J., Salgado, R., Santos, J., & Morais, M. (2020). Benthic diatom community
- dynamics in Mediterranean intermittent streams: Effects of water availability and their

- potential as indicators of dry-phase ecological status. *Science of the Total Environment*,
 719. <u>https://doi.org/10.1016/j.scitotenv.2020.137462</u>
- 11.37 Robson, B. J., Matthews, T. G., Lind, P. R., & Thomas, N. A. (2008). Pathways
 for algal recolonization in seasonally-flowing streams. *Freshwater Biology*, 53(12),
 2385–2401. <u>https://doi.org/10.1111/j.1365-2427.2008.02061.x</u>
- 11.38 Morón, M. C. (2021). Protein hydration shell formation: Dynamics of water in
 biological systems exhibiting nanoscopic cavities. *Journal of Molecular Liquids*, 337.
 https://doi.org/10.1016/j.molliq.2021.116584
- 805 11.39 Hostetter, H. P., & Hoshaw, R. W. (1970). Environmental Factors Affecting
 806 Resistance to Desiccation in the Diatom Stauroneis anceps. *American Journal of Botany*,
 807 57(5), 512–518. <u>http://www.jstor.org/stable/2441048</u>
- 11.40 Crowe, J. H., Carpenter, J. F., Crowe, L. M., & Anchordoguy, T. J. (1990). Are
 Freezing and Dehydration Similar Stress Vectors? A Comparison of Modes of Interaction
 of Stabilizing Solutes with Biomolecules'. In *CRYOBIOLOGY* (Vol. 27).
- 811 11.41 Smirnoff, N. (1993). The role of active oxygen in the response of plants to water
 812 deficit and desiccation. *New Phytologist*, *125*, 27–58.
- 813 11.42 Welsh, D. T. (2000). Ecological significance of compatible solute accumulation
 814 by micro-organisms: from single cells to global climate. *FEMS Microbiology Reviews*,
 815 24(3), 263–290. https://doi.org/10.1111/j.1574-6976.2000.tb00542.x
- 816 11.43 Passy, S.I. (2007). Diatom ecological guilds display distinct and predictable
 817 behavior along nutrient and disturbance gradients in running waters. *Aquat. Bot.* 86, 171–
 818 178. <u>https://doi.org/10.1016/j.aquabot.2006.09.018</u>
- 819 11.44 Evans, J. H. (1959). The Survival of Freshwater Algae During Dry Periods: Part
 820 II. Drying Experiments: Part III. Stratification of Algae in Pond Margin Litter and Mud.
 821 In *Source: Journal of Ecology* (Vol. 47, Issue 1).
- 822 11.45 Peterson, C. G. (1987). Influences of Flow Regime on Development and
 823 Desiccation Response of Lotic Diatom communities *Ecology*, 68 (4), 946–954.
 824 https://doi.org/10.2307/1938366
- 11.46 Elias, C. L., Calapez, A. R., Almeida, S. F. P., & Feio, M. J. (2015). From
 perennial to temporary streams: An extreme drought as a driving force of freshwater
 communities' traits. *Marine and Freshwater Research*, 66(5), 469–480.
 https://doi.org/10.1071/MF13312
- 829 11.47 Brooks, R. T. (2009). Potential impacts of global climate change on the hydrology
 830 and ecology of ephemeral freshwater systems of the forests of the northeastern United
 831 States. *Climatic Change*, 95(3–4), 469–483. <u>https://doi.org/10.1007/s10584-008-9531-9</u>
- 832 11.48 Butterwick, C., Heaney, S. I., & Talling, J. F. (2005). Diversity in the influence
 833 of temperature on the growth rates of freshwater algae, and its ecological relevance.
 834 *Freshwater Biology*, 50(2), 291–300. <u>https://doi.org/10.1111/j.1365-2427.2004.01317.x</u>
- 11.49 Lund, J. W. G. (1949). Studies on Asterionella: I. The Origin and Nature of the
 Cells Producing Seasonal Maxima. *Source: Journal of Ecology*, *37*(2), 389–419.

- 837 11.50 Suzuki Y., & Takahashi, M. (1995). Growth responses of several diatom species
 838 isolated from various environments to temperature. J. Phycol., 31, 880–888.
- 839 11.51 Souffreau, C., Vanormelingen, P., Verleyen, E., Sabbe, K., & Vyverman, W.
 840 (2010). Tolerance of benthic diatoms from temperate aquatic and terrestrial habitats to
 841 experimental desiccation and temperature stress. *Phycologia*, 49(4), 309–324.
 842 <u>https://doi.org/10.2216/09-30.1</u>
- Rousch, J. M., Bingham, S. E., & Sommerfeld, M. R. (2004). Protein expression
 during heat stress in thermo-intolerant and thermo-tolerant diatoms. *Journal of Experimental Marine Biology and Ecology*, 306(2), 231–243.
 https://doi.org/10.1016/j.jembe.2004.01.009
- 11.53 Lai, Y.-K., Li, C.-W., Hu, C.-H., & Lee, M.-L. (1988). Quantitative and
 qualitative analyses of protein synthesis during heat shock in the marine diatom Nitzschia
 alba (Bacillariophyceae). *Journal of Phycology*, 24, 509–514.
 https://doi.org/10.1111/j.1529-8817.1988.tb04255.x
- 11.54 Scoccianti V., Penna A., Penna N., & Magnani M. (1995). Effect of heat stress on
 polyamine content and protein pattern in Skeletonema costatum. *Marine Biology*, *121*,
 549–554.
- 854 11.55 Rousch, J. M., Bingham, S. E., & Sommerfeld, M. R. (2003). Changes in fatty
 855 acid profiles of thermo-intolerant and thermo-tolerant marine diatoms during temperature
 856 stress. *Journal of Experimental Marine Biology and Ecology*, 295(2), 145–156.
 857 https://doi.org/10.1016/S0022-0981(03)00293-4
- 11.56 Pörtner, H.-O., D.C. Roberts, H. Adams, I. Adelekan, C. Adler, R. Adrian, P. 858 Aldunce, E. Ali, R. Ara Begum, B. Bednar-Friedl, R. Bezner Kerr, R. Biesbroek, J. 859 Birkmann, K. Bowen, M.A. Caretta, J. Carnicer, E. Castellanos, T.S. Cheong, W. Chow, 860 G. Cissé, S. Clayton, A. Constable, S.R. Cooley, M.J. Costello, M. Craig, W. Cramer, R. 861 Dawson, D. Dodman, J. Efitre, M. Garschagen, E.A. Gilmore, B.C. Glavovic, D. Gutzler, 862 M. Haasnoot, S. Harper, T. Hasegawa, B. Hayward, J.A. Hicke, Y. Hirabayashi, C. 863 Huang, K. Kalaba, W. Kiessling, A. Kitoh, R. Lasco, J. Lawrence, M.F. Lemos, R. 864 Lempert, C. Lennard, D. Ley, T. Lissner, Q. Liu, E. Liwenga, S. Lluch-Cota, S. Löschke, 865 S. Lucatello, Y. Luo, B. Mackey, K. Mintenbeck, A. Mirzabaev, V. Möller, M. 866 Moncassim Vale, M.D. Morecroft, L. Mortsch, A. Mukherji, T. Mustonen, M. Mycoo, J. 867 Nalau, M. New, A. Okem, J.P. Ometto, B. O'Neill, R. Pandey, C. Parmesan, M. Pelling, 868 P.F. Pinho, J. Pinnegar, E.S. Poloczanska, A. Prakash, B. Preston, M.-F. Racault, D. 869 870 Reckien, A. Revi, S.K. Rose, E.L.F. Schipper, D.N. Schmidt, D. Schoeman, R. Shaw, 871 N.P. Simpson, C. Singh, W. Solecki, L. Stringer, E. Totin, C.H. Trisos, Y. Trisurat, M. van Aalst, D. Viner, M. Wairiu, R. Warren, P. Wester, D. Wrathall, and Z. Zaiton Ibrahim, 872 873 2022: Technical Summary. [H.-O. Pörtner, D.C. Roberts, E.S. Poloczanska, K. Mintenbeck, M. Tignor, A. Alegría, M. Craig, S. Langsdorf, S. Löschke, V. Möller, A. 874 Okem (eds.)]. In: Climate Change 2022: Impacts, Adaptation and Vulnerability. 875 Contribution of Working Group II to the Sixth Assessment Report of the 876 Intergovernmental Panel on Climate Change [H.-O. Pörtner, D.C. Roberts, M. Tignor, 877 E.S. Poloczanska, K. Mintenbeck, A. Alegría, M. Craig, S. Langsdorf, S. Löschke, V. 878

- Möller, A. Okem, B. Rama (eds.)]. Cambridge University Press, Cambridge, UK and
 New York, NY, USA, pp. 37–118, <u>https://doi:10.1017/9781009325844.002</u>
- 11.57 Buma, A. G. J., Zemmelink, H. J., Sjollema, K., & Gieskes, W. W. C. (1996).
 UVB radiation modifies protein and photosynthetic pigment content,volume and ultrastructure of marine diatoms. *Marine Ecology Progress Series*, 142(1–3), 47–54.
 https://doi.org/10.3354/meps142047
- 11.58 Vincent, W. F., & Neale, P. J. (2009). Mechanisms of UV damage to aquatic
 organisms. In *The Effects of UV Radiation in the Marine Environment* (pp. 149–176).
 Cambridge University Press. <u>https://doi.org/10.1017/cbo9780511535444.007</u>
- 11.59 Häder, D. P., Williamson, C. E., Wängberg, S. Å., Rautio, M., Rose, K. C., Gao,
 K., Helbling, E. W., Sinha, R. P., & Worrest, R. (2015). Effects of UV radiation on aquatic
 ecosystems and interactions with other environmental factors. In *Photochemical and Photobiological Sciences* (Vol. 14, Issue 1, pp. 108–126). Royal Society of Chemistry.
 <u>https://doi.org/10.1039/c4pp90035a</u>
- 893 11.60 Ellegaard, M., Lenau, T., Lundholm, N., Maibohm, C., Friis, S. M. M., Rottwitt,
 894 K., & Su, Y. (2016). The fascinating diatom frustule—can it play a role for attenuation
 895 of UV radiation? In *Journal of Applied Phycology* (Vol. 28, Issue 6, pp. 3295–3306).
 896 Springer Netherlands. https://doi.org/10.1007/s10811-016-0893-5
- 897 11.61 De Tommasi, E., Congestri, R., Dardano, P., De Luca, A. C., Managò, S., Rea, I.,
 898 & De Stefano, M. (2018). UV-shielding and wavelength conversion by centric diatom
 899 nanopatterned frustules. *Scientific Reports*, 8(1). <u>https://doi.org/10.1038/s41598-018-</u>
 900 <u>34651-w</u>
- 11.62 Roy, S. (2009). Strategies for the minimisation of UV-induced damage. In *The Effects of UV Radiation in the Marine Environment* (pp. 177–205). Cambridge University
 Press. https://doi.org/10.1017/cbo9780511535444.008
- 11.63 Ha, S. Y., La, H. S., Min, J. O., Chung, K. H., Kang, S. H., & Shin, K. H. (2014).
 Photoprotective function of mycosporine-like amino acids in a bipolar diatom (Porosira glacialis): Evidence from ultraviolet radiation and stable isotope probing. *Diatom Research*, 29(4), 399–409. <u>https://doi.org/10.1080/0269249X.2014.894945</u>
- 11.64 Aguirre, L. E., Ouyang, L., Elfwing, A., Hedblom, M., Wulff, A., & Inganäs, O.
 (2018). Diatom frustules protect DNA from ultraviolet light. *Scientific Reports*, 8(1).
 https://doi.org/10.1038/s41598-018-21810-2
- 911 11.65 Wu, Y., Yue, F., Xu, J., & Beardall, J. (2017). Differential photosynthetic
 912 responses of marine planktonic and benthic diatoms to ultraviolet radiation under various
 913 temperature regimes. *Biogeosciences*, 14(22), 5029–5037. <u>https://doi.org/10.5194/bg-14914 <u>5029-2017</u>
 </u>
- 915 11.66 Falasco, E., Bona, F., Risso, A. M., & Piano, E. (2021). Hydrological
 916 intermittency drives diversity decline and functional homogenization in benthic diatom
 917 communities. Science of the Total Environment, 762.
 918 https://doi.org/10.1016/j.scitotenv.2020.143090

11.67 Leigh, C. & Datry, T. (2017). Drying as a primary hydrological determinant of
biodiversity in river systems: a broad-scale analysis', *Ecography*, vol. 40, no. 4, pp. 487–
499. https://doi.org/10.1111/ecog.02230

11.68 Tornés, E., & Ruhí, A. (2013). Flow intermittency decreases nestedness and
specialisation of diatom communities in Mediterranean rivers. *Freshwater Biology*,
58(12), 2555–2566. https://doi.org/10.1111/fwb.12232

- 11.69 B-Béres, V., Tóthmérész, B., Bácsi, I., Borics, G., Abonyi, A., Tapolczai, K.,
 Rimet, F., Bouchez, Á., Várbíró, G., & Török, P. (2019). Autumn drought drives
 functional diversity of benthic diatom assemblages of continental intermittent streams. *Advances* in Water Resources, 126, 129–136.
 https://doi.org/10.1016/j.advwatres.2019.02.010
- 930 11.70 Nemes-Kókai, Z., Kovács, K., Borics, G., Mayer, R., Novák, Z., Robotka, A. G., József, J., Érczes, K., Lukács, A., & B-Béres, V. (2023). Continuous precipitation loss 931 induced more pronounced compositional and diversity changes in the lotic phytobenthos 932 933 one-off drought events. Ecol. Indic., 148, 110051. than https://doi.org/10.1016/j.ecolind.2023.110051 934
- 11.71 Cantonati, M., Kelly, M. G., Demartini, D., Angeli, N., Dörflinger, G., 935 Papatheodoulou, A., & Armanini, D. G. (2020). Overwhelming role of hydrology-related 936 variables and river types in driving diatom species distribution and community 937 938 assemblage in streams in Cyprus. Ecological Indicators, 117. https://doi.org/10.1016/j.ecolind.2020.106690 939
- 11.72 Falasco, E., Piano, E., & Bona, F. (2016). Diatom flora in Mediterranean streams:
 flow intermittency threatens endangered species. *Biodiversity and Conservation*, 25(14),
 2965–2986. <u>https://doi.org/10.1007/s10531-016-1213-8</u>
- 11.73 Kelly, M. G., King, L., Jones, R. I., Barker, P. A., & Jamieson, B. J. (2008).
 Validation of diatoms as proxies for phytobenthos when assessing ecological status in lakes. *Hydrobiologia*, 610(1), 125–129. <u>https://doi.org/10.1007/s10750-008-9427-8</u>
- 946 11.74 INAG I.P. (2008). Manual para a avaliação biológica da qualidade da água em
 947 sistemas fluviais segundo a Directiva-Quadro da Água. Protocolo de amostragem e
 948 análise para o Fitobentos-Diatomáceas.
- 949 11.75 European Committee for Standardization (2003). European Standard EN
 950 13946:2003. Water quality Guidance standard for the routine sampling and pretreatment
 951 of benthic diatoms from rivers.
- 11.76 Stubbington, R., Paillex, A., England, J., Barthès, A., Bouchez, A., Rimet, F.,
 Sánchez-Montoya, M. M., Westwood, C. G., & Datry, T. (2019). A comparison of biotic
 groups as dry-phase indicators of ecological quality in intermittent rivers and ephemeral
 streams. *Ecological Indicators*, 97, 165–174.
 https://doi.org/10.1016/j.ecolind.2018.09.061
- 957 11.77 Barthés, A., Leflaive, J., Coulon, S., Peres, F., Rols, J.-L., Ten-Hage, L., 2015.
- 958 Impact of drought on diatom communities and the consequences for the use of diatom

- 959 index values in the river Maureillas (Pyrénées-Orientales, France). *River Res. Appl.*, 31,
 960 993–1002. <u>https://doi.org/10.1002/rra.2793</u>
- 961 11.78 B-Béres V., Kókai Z., Várbiró G., Mustazhapova G., Csabai, Z., Pernecker B.,
 962 Borics, G., Bácsi, I., Boda, P. (2022). Flow Intermittence Drives the Benthic Algal
 963 Composition, Biodiversity and Diatom-Based Quality of Small Hilly Streams in the
 964 Pannonian Ecoregion, Hungary, *Front Ecol Evol*, vol. 10,
- 965 <u>https://doi.org/10.3389/fevo.2022.834548</u>