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Abnormal regulation of pancreatic β-cell Na,K-ATPase on glucose intolerant rats

Author Block: A.R. Costa^{1,2}, C.M. Antunes^{1,3}, J. Cruz-Morais^{1,2};

¹Chemistry, University of Évora, Évora, Portugal, ²ICAAM - Institute of Mediterranean Agricultural and Environmental Sciences, Évora, Portugal, ³CNC - Centre for Neurosciences and Cell Biology, Coimbra, Portugal.

Abstract:

Background and aims: Glucose (G) is the most important physiological insulin secretagogue. It is widely accepted that, in pancreatic -cell, G evoked early ionic events such as membrane depolarization and Ca²⁺ influx through voltage dependent Ca²⁺ channels triggers insulin exocytosis. However, the role of other electrogenic systems, namely ionic pumps, to these events remains essentially uninvestigated. It is known that the activity of Na,K-ATPase is modified in type 2 diabetes (T2D). The pump is responsible for maintaining Na⁺ and K⁺ gradients across the plasma membrane and generates a net outward current as a result of 3Na⁺/2K⁺ exchange. It remains elusive whether Na,K-ATPase activity is regulated by G in pancreatic β-cell and/or this current contributes to the ionic events regulating insulin secretion. The aim of this work was to assess G evoked regulation of Na,K-ATPase activity in intact -cells of normal and G intolerant rats. Materials and methods: Pancreatic -cells, from normal (controls) or glucose-intolerant Wistar rats (GIR), were isolated and cultured (48h). Cell batches were pre-incubated (30min) with 2.1mM G to reach basal. Afterwards cells were challenged with [G] in the interval 0-11.1mM for 60min, for dose-dependence evaluation, or with 8.4mM G for 5-120min, for time-dependence evaluation. ATPase activity was assessed in intact cells by colorimetric quantification of Pi formed in 30min. Na,K-ATPase activity was calculated by the difference between the activities obtained in the absence and in presence the of 1mM ouabain. Results: G evoked both time- and dose-dependent regulation of Na,K-ATPase. In β-cells from controls, G induced a bimodal regulation of Na,K-ATPase. In the absence of G, Na,K-ATPase activity was 0.056±0.015U/mg. Raising [G] to 2.1mM induced a ≈3 fold increase of Na,K-ATPase activity whereas a further increase in [G] in the interval of 5.6-11.1mM evoked a significant reduction of Na,K-ATPase activity to the levels observed in the absence of the secretagogue. Compared to 2mM G, the activity was reduced in 68%, 55% and 66% when [G] was increased to 5.6, 8.4 and 11.1mM, respectively (n=3-12). GIR β -cells exhibit an altered profile of response to the secretagogue; In the absence of G, Na,K-ATPase activity was ≈4 fold the activity observed in the controls (0.202±0.036U/mg; n=3). The pump activity remained unchanged for 2.1-5.6mM G and similar to maximal activity observed in the controls (0.188±0.035 U/mg, for 2.1mM G; n=4). A significant reduction of the pump activity in GIR β -cells was induced by 8.4mM G (0.118±0.018 U/mg). G (8mM) induced a time-dependent inhibition of Na,K-ATPase with a biphasic profile. Pump activity decreased to a minimum value (32%) after 20min exposure to G, showing a partial recovery to 45%, 46% and 47% for 30, 60 and 120min, respectively (n=5-12). GIR β -cells showed an attenuated response to G (59% activity after 20min) without any recovery (n=5-11).

Conclusions: This work demonstrates that Na,K-ATPase is finely regulated by G in pancreatic β -cell from normal subjects. This regulation is impaired in GIR where desensitization and an attenuation of the inhibitory action of G were observed. In summary, Na,K-ATPase contribution to G-induced ionic events and insulin secretion might be relevant in T2D development.

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European Association for the Study of Diabetes (EASD) Rheindorfer Weg 3 D-40591 Düsseldorf - Germany Tel: +49-211-758 469 0 - Fax: +49-211-758 469 29 Web: <u>http://www.easd.org</u> E-mail: <u>abstracts@easd.org</u>

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