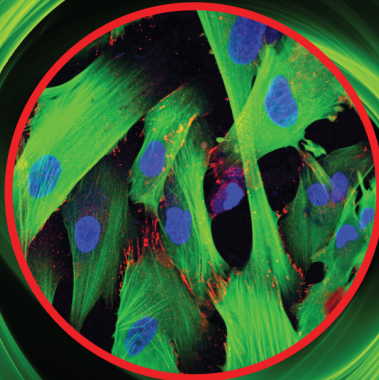


CELL BIOLOGY RESEARCH PROGRESS

# Cadherins

*Types, Structure and Functions*



Jonathan McWilliam  
Editor

NOVA

**CELL BIOLOGY RESEARCH PROGRESS**

# **CADHERINS**

## **TYPES, STRUCTURE AND FUNCTIONS**

**JONATHAN MCWILLIAM**

**EDITOR**



*Chapter 3*

**DISRUPTION OF E-CADHERIN PATTERN IN  
UTERINE AND MAMMARY TUMOURS**

*Adelina Gama<sup>1,2</sup>, Fernanda Seixas<sup>1,2</sup>,  
Maria dos Anjos Pires<sup>1,2,\*</sup>, Fernando Schmitt<sup>3,4</sup>  
and Rita Payan-Carreira<sup>5</sup>*

<sup>1</sup>CECAV, Universidade de Trás-os-Montes e Alto Douro (UTAD),  
Vila Real, Portugal

<sup>2</sup>ECAV, Dept. Veterinary Sciences, Universidade de  
Trás-os-Montes e Alto Douro (UTAD), Vila Real, Portugal

<sup>3</sup>IPATIMUP, Institute of Molecular Pathology and Immunology of the  
University of Porto, Porto, Portugal

<sup>4</sup>Faculty of Medicine of the University of Porto, Porto, Portugal

<sup>5</sup>MED - Mediterranean Institute for Agriculture, Environment and  
Development & Dept. of Veterinary Medicine, ECT,  
Universidade de Évora [Pole at Mitra], Évora, Portugal

---

\* Corresponding Author's E-mail: apires@utad.pt.

## ABSTRACT

E-cadherin (E-cadh), a member of the classic cadherins superfamily, plays an important role in epithelial cell-to-cell adhesion, encompassing the dynamic interactions between adjacent cells including the control of morphogenesis, maintenance of cell polarity and tissue architecture. Cadherins comprise a large family of cell surface glycoproteins, presenting unique extracellular regions domains known as the cadherin motifs or domains, which fold like immunoglobulin domains. They mediate strong  $\text{Ca}^{2+}$ -dependent homophilic interactions between neighbouring epithelial cells, resulting in the formation of cell adhesion "zippers." E-cadh cytoplasmic tail links to catenins and, thereby to the actin cytoskeleton and signalling proteins to form a cell-cell signalling centre: it regulates several intracellular signal transduction pathways, including Wnt/ $\beta$ -catenin, PI3K/Akt, Rho GTPase, and NF-KB signalling.

E-cadh plays a crucial role in the barrier formation of polarized epithelial cell layers at the interfaces contacting with the external environment, namely the uterus and the mammary gland. The maintenance of these barriers could be considered as a prime immunologic function of E-cadh, compartmentalizing potentially harmful agents away from the underlying tissue. Disruption of classical cadherin expression has been related to the occurrence of diseases driving disturbances in tissue architecture, such as inflammation and cancer.

In cancer, loss of E-cadh expression/function increases cell proliferation, cell migration, and disruption of epithelial cell homeostasis, driving cell dissociation and scattering. Alterations of E-cadh expression have also been reported during particular moments of the female reproductive physiology, namely throughout the oestrous cycle or during the embryo-maternal interaction at embryo implantation (early pregnancy). Several studies have shown the downregulation of E-cadh in malignant epithelial tumours, which has been associated with loss of cell differentiation, epithelial to mesenchymal transition (EMT) and invasion. Data also suggest that loss of E-cadh may be associated with malignant progression, metastasis, and reduced survival in multiple cancer patients.

In this chapter, we review and discuss the role of E-cadh in the uterine and mammary gland homeostasis and describe disruptive patterns of E-cadh expression in neoplastic conditions of the uterus and mammary glands in human and domestic dogs and cats.

**Keywords:** E-cadherin, uterus, mammary gland, carcinoma, human, dog, cat

## **INTRODUCTION**

Cadherins are cell surface glycoproteins generally associated with cell-to-cell adhesion and recognition processes occurring in animal tissues. Cadherins form a superfamily of more than one hundred of calcium-dependent membrane proteins mediating homophilic cell-to-cell adhesion [1]. Presenting unique extracellular regions domains, known as the cadherin motifs or cadherin domains, which fold like immunoglobulin domains [2], these proteins have been found in both invertebrates and vertebrates, and in a wide array of tissues [3, 4].

Yet, classic cadherin roles go far beyond the epithelial cell-to-cell adhesion. They control cell movements underlying morphogenesis, changes in cell polarity, cell structure, and also mediate several intracellular signalling processes associated with cytoplasmic organization and motile behaviours of cells, as well as changes in gene expression to control cell differentiation and growth, and tissue architecture [5, 6]. Cadherins contribute to tissue homeostasis, participating in the tissue barrier function, cell proliferation, and migration [7]. As other type I classic cadherins, E-cadherin (E-cadh) mediates cell adhesion and is a key determinant of epithelial morphology and differentiation in most epithelial body tissues and organs [8].

### **E-Cadherin at the Core of Epithelial Adherens Junctions**

Epithelial integrity depends on the interaction of different types of junctions, namely the tight junctions, adherens junctions and desmosomes. Together, they constitute the epithelial junctional complex. Adherens junctions (AJs) are specialized cell-to-cell adhesion sites consisting of E-cadh/catenin complexes that link to actin cytoskeleton [6], which in turn regulates the assembling, organization, stability and remodeling of AJs [9]. Connection with actin bundles allows the interaction of other types of junctions with AJs, working together to maintain the epithelial barrier. In the case of AJs, the barrier is achieved by creating a continuous adhesive

belt at the apical–lateral interface of adjacent cells. Despite providing strength and polarity to epithelial barriers, AJs present notable plasticity, which is important for tissue morphology and morphogenesis [10]. This plasticity is of utmost importance to coordinated multicellular movement or the cooperatively regulated single-cell migration [11, 12], either for embryo morphogenesis, tissue healing and tumour invasion/progression.

As said, clusters of dimeric E-cadh mediate trans-homophilic interactions between neighboring cells forming AJs [10, 13]. The plasticity of the epithelial barrier is made possible by a rapid and constant turnover of membrane E-cadh, driving its internalization through different endocytic pathways and its recycling by exocytosis [13]; in contrast, the *de novo* synthesis of E-cadh is relatively slow. The E-cadh internalization process is mediated by clathrin, which allows the phenomenon to be spacially controlled [10].

Inside the epithelial cells, the E-cadh cytoplasmic domain links to  $\beta$ -catenin and p120-catenin; via  $\beta$ -catenin, E-cadh links to  $\alpha$ -catenin and thereby to actin [9, 13, 14].  $\alpha$ -catenin links to actin through either the F-actin or vinculin [13]. The binding of  $\alpha$ -catenin to F-actin provides stability to the adhesion point, while p120-catenin stabilises the cell-to-cell adhesion by controlling the retention of E-cadh at the cell surface [9]. While linked to p120-catenin, E-cadh is guarded from clathrin-mediated endocytosis, driving the notion that p120-catenin acts as a master regulator of cadherin stability [10, 12]. The fixation of E-cadh in the cell membrane results from the inhibition of endocytosis [13]. Besides its control of cadherin internalization, p120-catenin is considered as a “set-point” for cadherin expression [15]. It is now accepted that the level of E-cadherin expression, not that of the catenins, is the rate-limiting step for the formation of E-cadh complexes and the cell adhesion [16]. It has been proposed that p120-catenin regulates the cadherin levels within the cell, and possibly the switching of the pattern of cadherin expression from one member to another, specially in cells expressing multiple cadherin types. Moreover, it has been shown that p120-catenin may shuttle to the nucleus, where it interacts with the transcription factor Kaiso, which is involved with the regulation of various cancer-related genes [17].

Depletion of extracellular calcium drives the disruption of adhesion between adjacent epithelial cells [10]. Remodelling of AJs implies the control of E-cadh internalization, and thereby the disturbance of the E-cadh/catenin complexes. The disruption of these complexes is regulated mainly by protein kinases and phosphatases that phosphorylate the AJs structural proteins [13], affecting the connection of E-cadh to the actin cytoskeleton. Small GTPases, such as the RhoGTPases, Rac and Cdc42, and corresponding effectors, have been associated to AJs remodelling [9, 10, 13]. Furthermore, it has also been proposed that Src family of non-receptors protein kinases (e.g., c-Src and c-Yes) are essential players in AJs remodelling [18]. The activation of Src kinases drives the phosphorylation of AJ components and the subsequent disruption of cell-to-cell adhesions, therefore contributing to cell migration.

### **E-Cadherin as a Key Modulator during Development and Neoplastic Progression**

Cadherin endocytosis plays a significant role in physiological and pathological processes. Cadherin endocytosis is crucial in the embryo differentiation and development [5], as well as in canine trophoblast migration at implantation [19]. A similar event occurs during epithelial-mesenchymal transition (EMT), where loss of membrane E-cadh (either by cadherin endocytosis or cadherin switching) favour proliferation, migration and invasion of neoplastic cells [15, 20].

Cadherin internalization may occur through different endocytic pathways, the clathrins' being the most studied [10, 15]. Additional pathways associated to cadherin internalization include the caveolin-mediated and macropinocytosis-like pathways. An increase in the phosphorylation of cadherin/catenin complexes drives a loss of interaction with the actin cytoskeleton, the internalization of membrane cadherin, and the loss of intercellular adhesion [21]. Nevertheless, not all the molecules entering an endocytic pathway undergo lysosomal degradation. Some cadherin molecules are recycled back to the cell membrane, allowing cells

to retain their barrier properties and their polarity. The fate of E-cadh trafficking within the cell (degradation *vs.* recycling) is influenced by a family of Src non-receptors protein kinases [16]. c-Src and c-Yes seem to have opposing roles in the outcome of the complex disassembled proteins: c-Yes drives the molecules towards the endocytic vesicles trafficking mechanism (recycling back to the membrane), whereas c-Src fosters their lysosomal degradation [16, 22]. In some tumours, Src induced down-regulation of E-cadherin depends on integrin signaling and FAK phosphorylation [23]. Increased neoplastic Src activity have been associated with tumour invasiveness [16] and the loss of membrane E-cadh expression which suppresses cell-to-cell contacts and favours cell migration, invasiveness and metastatic dissemination. Therefore, E-cadh is often used as a prognostic marker for several solid tumours [16, 24].

Dysregulation of the E-cadh complexes may drive cells into the transition to a different cell phenotype, contributing to EMT which is often associated with tumour progression and metastases. During EMT, epithelial cells lose polarity and the typical intercellular adhesion, and develop mesenchymal features together with increased migratory and invasive properties [20]. However, EMT does not occur only in cancer; it can also be observed in natural biological phenomena such as during embryogenesis, wound healing [25], or even during glandular cyclic branching (e.g, the branching of the mammary ducts) [26].

Although initial studies pointed to a reductionist binary process, EMT is now proposed as a highly coordinated plastic program, manifesting as dynamic transitional states between the epithelial and mesenchymal phenotypes [27, 28], with the identification of distinct intermediate states (epithelial, intermediate hybrid and mesenchymal states) [27, 29].

EMT is not a mandatory pre-requisite for tumour cell invasion and metastasis [23, 30]. In fact, neoplastic cells can move in a collective manner, on which depend on the disturbance of cadherin-mediated cell-to-cell contacts. This invasive behaviour is also determined by p120-catenin, and may be accomplished by either an E-cadh-dependent or independent mechanism. Down-regulation of E-cadh can be achieved by the regulation of Rac and Cdc42 activity or indirectly by inhibiting RhoGTPases and



downstream cytoskeletal dynamics involved in cell invasion and migration [31].

Cadherin switching, that alters intercellular adherence and the cadherin-associated signaling pathways, is a common event during EMT. While normal epithelial cells express particular patterns of cadherins, cells undergoing EMT start expressing different cadherin types (mesenchymal cadherins), such as Neuronal (N-) cadh, Placental (P-) Cadh, or cadherin 11, among others [20]. This switch seldom occurs at once, but follows a specific pattern, possibly responding to different internal or external stimuli. TGF- $\beta$  and the Snail, Slug and Twist transcription factors have been proposed as main EMT regulators. Dysregulation of EMT may ultimately drive to cancer [25]. Cadherin switch may coordinate changes in several cell functions, namely in cell metabolism, resistance to hypoxia and programmed cell death mechanisms, and lower adhesion to extracellular matrices [32], which all contribute to proliferation and the invasive behaviour of neoplastic cells.

The loss of functional E-cadh will contribute to tumour cells resistance to apoptosis mediated by either the intrinsic or the extrinsic pathways [33-36]. More recently, Capra and Eskelinen [37] showed that the disruption of E-cadh mediated adhesion triggers the up-regulation of survivin, which suppresses apoptosis and promotes cell proliferation and migration; survivin signalling may therefore play an important role in neoplastic progression in cells not suffering EMT.

## **E-CADHERIN EXPRESSION IN UTERINE CANCER**

### **E-Cadherin in the Normal Endometrium**

The cadherin/catenin complex is an essential intercellular adhesive system in the mammal endometrium, where it coordinates key morphogenetic processes, regulates epithelial differentiation and proliferation, and supports the epithelial phenotype [38]. Moreover, it also

mediates the interactions between the embryo and maternal tissues at implantation [19, 39, 40].

The few available studies on the expression of E-cadherin and/or  $\beta$ -catenin have established the existence of cyclic variations of these molecules throughout the uterine cycle and early pregnancy in humans [41-43], sheep [44], pigs [45] and dogs [19, 46].

Albeit studies on gene expression and protein location or quantification are not always concordant, it has been proposed that progesterone determines a decline in the strength of AJs which would facilitate the trophoblast invasion across the epithelial barrier [47]. This hypothesis is supported by a decrease in E-cadherin around the implantation time in ewes [44], sows [48], and dogs [19]. In the feline cyclic endometrium, a similar decrease in the intensity of E-cadherin membrane labelling has been found during progesterone dominance (M.A. Pires, unpublished results). Dudley et al. [49] described a dislocalization of E-cadherin from the lateral plasma membrane to the cytoplasm in uterine epithelial cells in early pregnant cats.

Furthermore, studying the canine embryo-maternal interactions at implantation Payan-Carreira et al. [19] showed that internalization of E-cadherin occurs in the maternal superficial epithelial layers of the endometrium, while the invading trophoblast retain the membrane labelling. This work agrees with an involvement of E-cadherin/catenin complex in the embryo implantation and further suggests that the adhesiveness strength favours the collective invasion of trophoblastic cells.

## **E-Cadherin in Endometrial Carcinomas**

The loss of cellular polarity [50] and alterations in cell adhesion [38] are hallmarks characteristics of cancer. Multiple studies reported reduced or aberrant expression of E-cadherin and/or catenins in different human epithelial cancers: thyroid and esophageal carcinoma [38], breast carcinoma [38, 51, 52], gastric and pancreatic carcinoma, bladder and prostatic carcinoma, among others [38]. Defects in the E-cadherin/catenin

adhesion complex have been described in endometriosis [43] and in several gynecologic carcinomas, including ovarian, endometrial [38, 52, 53] and cervical carcinomas [38]. Endometrial cancer is the most common malignancy of the women genital tract. In the majority of cases the prognosis is good, but women with poor differentiated, deep myoinvasive tumours, or with extension of disease to other organs or lymph nodes within the pelvis, have frequently disease recurrence [54].

In endometrial cancer, E-cadh is a recognized putative marker of good prognosis [52, 55]. Loss of E-cadh expression, has been shown to drive the loss of cell adhesiveness, and contact inhibition [52], promoting tumour progression and an aggressive behaviour, invasion and metastasis in several epithelial tumours [52, 56]. In endometrial cancer, it is correlated with tumour dedifferentiation [50, 57], high grade histology [50, 54, 56], and deep myometrial invasion [50, 54, 57], higher rate of extrapelvic recurrence [54], presentation of other adverse prognostic factors and lower overall survival [52, 56]. Some studies showed that E-cadh expression is an independent prognostic factor for women endometrial carcinoma [54, 55]. Decreased membranous E-cadh expression is predictive for endometrial cancer mortality, disease progression, and extrapelvic recurrence, independent of known prognostic factors such as stage, grade, and histological subtype [54].

Feline endometrial adenocarcinomas are uncommon, poorly characterized lesions [58], that may be underdiagnosed [59], affecting even young cats [60, 61]. There are few published studies on the immunophenotype of these lesions [58] and with small case series [59-61] which makes the full characterization of these neoplasms difficult. Though, some studies reported E-cadh and  $\beta$ -catenin expression in both the normal feline endometrium [58], and in endometrial adenocarcinoma [58, 62]. According to Gil da Costa et al. [58], the loss of cell adhesion that occurs within these tumours does not require down-regulation of E-cadh expression; in addition, the nuclear translocation of  $\beta$ -catenin was not a characteristic feature of feline endometrial carcinomas.

In their study, Carico et al. [38] showed that human endometrial carcinomas may present different patterns of E-cadh and  $\alpha$ -catenin

expression within the tumour, reflecting the intratumoral heterogeneity of the neoplastic epithelium. Similarly, feline endometrial carcinomas present a patchy pattern of the cadherin/catenin complexes within the tumour, presenting areas of membrane and areas of cytoplasmic E-cadh expression (M.A. Pires, unpublished results). It is possible that while remaining anchored to their neighbours due to E-cadh membranar expression, epithelial neoplastic cells might have an impaired ability to metastasize. This hypothesis deserves to be further explored in the case of feline endometrial carcinomas, where a benign clinical course has been frequently described [60, 63, 64].

Canine endometrial carcinomas are rare, and mostly occur in old bitches [65]. Despite both E-cadh and  $\beta$ -catenin expression have been reported in the normal cyclic endometrium and the early pregnant uterus [19], the available literature is sparse in studies of adhesion molecule expression on canine endometrial carcinomas. This gap on information on E-cadh and other adhesion molecules on promotion, progression and prognosis of feline and canine endometrial carcinomas shows the need of new studies on this topic to evaluate the real effect of adhesion molecules on uterus tumourigenesis.

## **E-CADHERIN EXPRESSION IN MAMMARY CANCER**

### **E-Cadherin Expression in Normal Mammary Tissues**

As stated above, E-cadh has been described in most epithelial tissues, including the mammary gland. In normal human breast and in canine and feline mammary gland, E-cadh is expressed by luminal epithelial cells at the cell membrane [66-69]; this pattern of expression is also observed for  $\alpha$ - and  $\beta$ -catenins [70-73]. In contrast, P-cadh is restricted to the basal myoepithelial cells [74-76]. Although cadherins expression in the mammary differentiated cells is well defined and cell-type specific, their expression in progenitor or mammary stem cell populations remain unclear [26]. Basal, P-cadh expressing progenitor cells seems to be responsible for

mammary formation and branching morphogenesis, while alveolar progenitor cells most likely represent luminal cells which give rise to E-cadh positive cells [26, 77]. It has been proven that E-cadh is essential for mammary gland, after conditional E-cadh inactivation studies using knockout mouse models [26, 78].

## **E-Cadherin Expression in Mammary Cancer**

Breast cancer is among the deadliest malignancies in developed countries [79], with the metastatic spread being the primary reason of this fatal outcome [79-81]. In canine species, spontaneous mammary tumours are the second most frequent tumour, and the most common neoplasia in the female dog. Regarding its biological behaviour, malignant cases account for up to 50% of female mammary tumours [82, 83]. In feline species, mammary neoplasias are among the most commonly diagnosed tumours in female cats [82, 84-86], accounting for 12% of all tumours and 17% of the tumours in queens [82]. Most studies reported that more than 80% of feline mammary tumours are malignant, along with rapid progression and metastasis [84, 86].

E-cadh, a putative tumour suppressor gene implicated in carcinogenesis [87, 88], is classically considered a good prognostic marker in cancer [89]; several studies proposed E-cadh as a tumour and invasion suppressor molecule, as invasion and metastasis are promoted when its expression is lost [70, 80, 90].

In human breast cancer, although representing one of the cancer types for which E-cadh has been extensively investigated for diagnostic and prognostic purposes, conclusions are inconsistent with regard to its relevance [89, 91-93]. Some studies associated low E-cadh expression with tumour size [93, 94], histological grade [93-95], distant metastasis and absence of oestrogen receptors, but not with lymph node status [94]. Although some investigators have found no association between E-cadh and the tumour stage, lymph node status or metastasis [89], other reported an association of E-cadh with lymph node status [93] and TNM stage [93].

With regard to prognosis, data is also controversial: many studies reported that E-cadh downregulation was associated with poor survival [80, 92-94, 96], but Gillett et al. [97] found that E-cadh reduction was a favorable prognostic factor, and Wang et al. [98] found no relationship between E-cadh and prognosis. These contrasting results may be associated to inter-study heterogeneity with respect to clinical data collection, immunohistochemical staining and interpretation as well as statistical modeling, that affect studies results [96].

Adhesion molecules have been extensively studied in canine mammary tumours [66-68, 70, 71, 83, 99-104]. In these tumours, E-cadh immunoexpression was first described by Restucci and coworkers [66], reporting low expression in malignant neoplasia. After, multiple studies showed reduced membranous expression of E-cadh in malignant mammary tumours, suggesting that down-regulation of adhesion molecules is a common event in canine mammary tumours [68, 71, 100]. Some studies associated low membrane expression of E-cadh and the histological type, poor differentiation, invasiveness, high proliferation and lymph node metastasis [66, 68, 71, 99, 100]. Reduction of E-cadh expression and associated catenins was also related with invasion and metastases on canine mammary tumours [66, 67, 70, 99]. These data hint a possible role of E-cadh in canine mammary tumours aggressiveness and on the emergence of an invasive and metastatic phenotype, suggesting E-cadh as a potential prognostic marker [67, 68, 71, 105]. However, although some studies found association between E-cadh and  $\beta$ -catenin expression and survival [68], other failed to find these associations [70, 100].

Regarding the feline mammary tumours, few studies have evaluated adhesion molecules expression and their prognostic value [106-108]. Loss of membrane E-cadh expression and its dislocation to the cytoplasm has been described in feline mammary carcinomas compared with benign tumours and hyperplastic or normal mammary tissues [106, 108, 109], as it was reported in canine mammary tumours [66, 68, 71, 100]. Albeit Penafiel-Verdu et al. [108] found an association between E-cadh expression and the grade and existence of regional metastasis at the moment of diagnosis, other studies fail to establish such associations [69,

109]. Abnormal cytoplasm E-cadh and/or catenin has been described in feline cancer cells [107-109] but not in the normal mammary tissue [107, 108], suggesting that this abnormal location might be related to a malignant transformation of feline mammary tumours [107]. Albeit not fully explored the prognostic role of E-cadh in feline mammary tumours, some studies could not establish an association between E-cadh and survival [109].

Understanding the metastatic progression of breast cancer in humans offers the opportunity to tailor individual based therapies by considering specific tumour characteristics [80]. For metastases to occur, several progressive changes are needed; these include neovascularization, decreased adherence of the tumour cells to each other, increased motility, adhesion to the extracellular matrix, and degradation of the extracellular matrix [80]. Downregulated expression of E-cadh destroys cell junctions and thus epithelial cells acquire the ability to migrate [93]. Consequently, decreased expression of E-cadh facilitates tumour invasion and metastasis [87, 88, 93]. Nevertheless, in the vast majority of human breast cancers, E-cadh expression is not lost, but retained [93]. Some studies in human reported preserved E-cadh expression in high aggressive breast cancer [110], such as the inflammatory breast cancer, in which E-cadh is not only retained but overexpressed [90, 95, 111] and distributed circumferentially 360° around the cancer cell membrane [93]. E-cadh accumulation and, subsequently, overexpression is responsible for the formation of lymphovascular emboli, conferring resistance to apoptosis and a survival advantage. It can be argued that, in the setting of the lymphovascular tumoral embolus, E-cadh is functioning not as a suppressor gene but rather as an oncogene [90]. More recently, Chu et al. [112] showed that E-cadh expression plays an important role in regulating tumourigenicity and hypoxia responses in an inflammatory breast cancer model: the loss of E-cadh and/or overexpression of its repressors, such as ZEB1, downregulated the expression of hypoxia-inducible 1 $\alpha$  transcription factor (HIF-1 $\alpha$ ), leading to a reduction of the extracellular acidification of inflammatory breast cancer, tumour growth and metastasis formation [113].

Although multiple studies reported that metastases were more common in women with breast cancer with absent or low E-cadh expression [91, 114], recent studies have called into question whether cancer cells require the loss of E-cadh to invade and metastasize [89]. Besides, results on the expression of E-cadh in both primary tumour and lymph node metastasis are contradictory. Some studies reported lymph node metastasis with reduced or loss of E-cadh expression [115], whereas others reported overexpression [83, 116] or a similar expression to that observed in the primary tumour [83, 114]. E-cadh overexpression in canine mammary lymph node metastases was also reported by some studies [66, 70]. In the female cat, differences in E-cadh pattern of expression between primary tumors and regional metastases were described [69, 109], suggesting that during mammary carcinogenic progression, there is a dynamic and reversible modulation of the E-cadh complex [69]. E-cadh overexpression in metastases might be associated to their stabilization in the new environment, in order to adhere and to re-establish tissue architecture [116]. Although the mechanism behind this re-expression remains unclear [70], it has been suggested that E-cadh expression is not usually inactivated but its expression is dynamically modulated during the metastatic cascade. This plasticity is probably related to the flexibility of adhesion complexes, which might be temporarily downregulated or not expressed in primary tumours allowing cell detachment and invasion, and lately recovered in the metastatic site, thus favoring the survival and growth of metastatic cells [66, 116]. So, the loss of E-cadh expression might be a transient phenomenon that allows malignant cells to invade vascular channels and tissues; once in circulation, cancer cells re-express E-cadh, facilitating intercellular adhesion and enabling the formation of cohesive tumour emboli [95].

As described previously, cancer invasion and metastasis are, in fact, highly versatile processes, regulated at multiple levels. Recent studies indicate that cancer cells utilize two major migratory strategies: preserving intercellular cohesion as a collective, or as single-cell invasion into the surrounding stroma. The process of cancer cell individualization and acquisition of an invasive migratory phenotype commonly occurs within



the framework of EMT. During EMT, epithelial cells undergo major transcriptional and morphological transformations, resulting in the loss of their intercellular adhesions, and the acquisition of mesenchymal-like properties [117]. As EMT progresses, the transformed cells lose the junctional connections with their neighbours, disengage from the epithelial layer in which they originated, and express mesenchymal markers such as N-cadh, vimentin, and a multitude of specific transcription factors (e.g., Snail, Slug, Twist) [117, 118]. The acquired mesenchymal phenotype is manifested in enhanced migratory activity, extracellular matrix production, invasiveness, and elevated resistance to apoptosis [118, 119]. These changes enable the cells to enter into small vessels and disseminate to distant organs, where they form metastasis [117].

There is controversy about the corollary of the type of E-cadh inactivation (gene mutation or promoter hypermethylation/repression) and the aggressiveness of tumour cells [87]. The infiltrating lobular carcinoma of human breast is classically characterized by a loss of E-cadh immunostaining and intercellular cohesion that results from truncating mutations or epigenetic bi-allelic silencing of E-cadh gene [89, 120-122].

In some breast carcinomas, invasion and metastasis is promoted when E-cadh expression is lost by the promoter methylation or repression by Snail/Slug and other EMT mediators [87, 90]. EMT has been associated with the metastatic cascade in several types of carcinomas, including human breast carcinomas [123, 124]; Lombaerts et al. [87] suggest that E-cadh promoter methylation, but not mutational inactivation, is part of the EMT programme, resulting in increased invasiveness and tumourigenic capacity in breast cancer. The molecular events of this programme can be inferred from the differentially expressed genes and include genes from the TGF $\beta$  pathway, transcription factors involved in E-cadh regulation (i.e., ZFH1B, SNAI2, but not SNAI1, TWIST), annexins, AP1/2 transcription factors and members of the actin and intermediate filament cytoskeleton organization. Altered expression of these transcription factors seems to be also associated with an altered overexpression of transcriptional repressors of E-cadh in tumour cells; thus, considering that metastasis is facilitated by

EMT, the disturbance of this process might prevent breast cancer dissemination [87].

Recently, Raposo-Ferreira et al. [125] provided evidences that EMT plays an important role in the metastatic process of canine mammary carcinomas, describing a significantly high co-expression of E-Cadh<sup>+</sup>/vimentin<sup>+</sup> in primary mammary carcinomas, especially in high grade carcinomas, when compared to their paired metastases. This distinct expression pattern suggests that EMT is also a dynamic reversible process in canine mammary tumours [125]. In fact, accumulating evidence supports a phenotypic plasticity of metastatic cells that allows a reverse process at a secondary site, known as mesenchymal-epithelial transition (MET), that promotes a transition back into an epithelial phenotype which will allow secondary tumour growth [27].

A few studies investigated the expression of EMT-inducing transcription factors, such as Snail, Slug or Zeb, and their correlation with E-cadh in canine or feline mammary neoplasia. With regard to Snail, Im et al. [126] found no association between Snail and E-cadh expression, although Snail expression in canine mammary tumours was significantly correlated with aggressive clinicopathological features such as histological grade and lymphatic invasion. In contrast, Gamba et al. [127] reported a direct association between E-cadh downregulation and Snail up-regulation in canine invasive micropapillary mammary carcinoma; however, no significant correlation was found between E-cadh and Zeb2 in this histological type [128]. Pang et al. [129] demonstrated that EMT induction by TGF $\beta$  can enrich cancer cells with stem cell properties.

Classic EMT is usually defined by morphological changes combined with the loss of E-cadh. However, *in vitro* studies using human breast cancer cell lines showed that EMT might be possible without E-cadh loss [130]. Timmermans-Sprang and collaborators [131] described that P-cadh mutations are associated with an EMT phenotype in canine mammary cell lines with E-cadh expression. In fact, in human breast cancer, the presence of P-cadh in an E-cadh positive background can promote invasion [132].

Regarding feline mammary tumours, Buendia et al. [133] found a negative association between E-cadh and N-cadh expression and reported

an association between N-cadh and the tumour histological grade and regional metastasis; these authors suggested that N-cadh expression could be considered a sign of malignancy. Another study showed a co-expression of E-cadh and P-cadh in both primary and metastatic lesions and reported a large number of P-cadh positive feline mammary tumours suggesting that cadherins switching to P-cadh could play an important role in feline mammary tumourigenesis [134]. This hypothesis deserve additional investigation.

## CONCLUSION

E-cadh is a classic cadherin that plays an important role in epithelial cell-to-cell adhesion, with recognized functions in morphogenetic processes and in the maintenance of normal tissue architecture. Alterations of E-cadh expression/function have been reported in physiological processes such as throughout the oestrous cycle or during the embryo-maternal interaction at embryo implantation. In contrast, E-cadh loss or reduced expression/function has also been associated with disease, namely with tumourigenesis and malignant progression of carcinomas. Breast cancer is one of the most studied cancer types regarding E-cadh relevance in the tumourigenesis, tumour progression and prognosis. Although the significant number of *in vitro* and *in vivo* investigations in this subject, several questions are still controversial. There is no consensus as to consider E-cadh as a prognostic marker, and its importance during tumour progression differs across the available literature. Nevertheless, E-cadh loss has long been described as a hallmark of EMT, a dynamic process that is especially relevant for breast cancer neoplastic invasion, progression and metastasis formation. In the veterinary setting, although fewer studies are available, especially for the endometrial cancer, results seem similar to the ones described for humans. Future studies are welcome, aiming the factual relevance of E-cadh in the metastatic cascade, focusing in the complex tumour-microenvironment interactions, both at the primary tumour and at the metastatic niche.

## REFERENCES

- [1] Colás-Algora, N., and J. Millán. 2019. "How many cadherins do human endothelial cells express?" *Cell Mol Life Sci* 76 (7):1299-1317. doi: 10.1007/s00018-018-2991-9.
- [2] Pokutta, S., and W. I. Weis. 2007. "Structure and mechanism of cadherins and catenins in cell-cell contacts." *Annu Rev Cell Dev Biol* 23:237-61. doi: 10.1146/annurev.cellbio.22.010305.104241.
- [3] Nollet, F., P. Kools, and F. van Roy. 2000. "Phylogenetic analysis of the cadherin superfamily allows identification of six major subfamilies besides several solitary members." *J Mol Biol* 299 (3):551-72. doi: 10.1006/jmbi.2000.3777.
- [4] Oda, H., and M. Takeichi. 2011. "Evolution: structural and functional diversity of cadherin at the adherens junction." *J Cell Biol* 193 (7):1137-46. doi: 10.1083/jcb.201008173.
- [5] Halbleib, J. M., and W. J. Nelson. 2006. "Cadherins in development: cell adhesion, sorting, and tissue morphogenesis." *Genes Dev* 20 (23):3199-214. doi: 10.1101/gad.1486806.
- [6] Gumbiner, B. M. 2016. "Classical Cadherins." In *The Cadherin Superfamily: Key Regulators of Animal Development and Physiology*, edited by Shintaro T. Suzuki and Shinji Hirano, 41-69. Tokyo: Springer Japan.
- [7] Garcia, M. A., W. J. Nelson, and N. Chavez. 2018. "Cell-Cell Junctions Organize Structural and Signaling Networks." *Cold Spring Harb Perspect Biol* 10 (4). doi: 10.1101/cshperspect.a029181.
- [8] Braga, V. 2016. "Spatial integration of E-cadherin adhesion, signalling and the epithelial cytoskeleton." *Curr Opin Cell Biol* 42:138-145. doi: 10.1016/j.ceb.2016.07.006.
- [9] Gloushankova, N. A., S. N. Rubtsova, and I. Y. Zhitnyak. 2017. "Cadherin-mediated cell-cell interactions in normal and cancer cells." *Tissue Barriers* 5 (3):e1356900. doi: 10.1080/21688370.2017.1356900.

- [10] Brüser, L., and S. Bogdan. 2017. "Adherens Junctions on the Move—Membrane Trafficking of E-Cadherin." *Cold Spring Harb Perspect Biol* 9 (3). doi: 10.1101/cshperspect.a029140.
- [11] Bajpai, A., J. Tong, W. Qian, Y. Peng, and W. Chen. 2019. "The Interplay Between Cell-Cell and Cell-Matrix Forces Regulates Cell Migration Dynamics." *Biophys J* 117 (10):1795-1804. doi: 10.1016/j.bpj.2019.10.015.
- [12] Katsuno-Kambe, H., and A. S. Yap. 2020. "Endocytosis, cadherins and tissue dynamics." *Traffic* 21: 268–273. doi: 10.1111/tra.12721.
- [13] Takeichi, M. 2014. "Dynamic contacts: rearranging adherens junctions to drive epithelial remodelling." *Nat Rev Mol Cell Biol* 15 (6):397-410. doi: 10.1038/nrm3802.
- [14] Mège, R. M., and N. Ishiyama. 2017. "Integration of Cadherin Adhesion and Cytoskeleton at." *Cold Spring Harb Perspect Biol* 9 (5). doi: 10.1101/cshperspect.a028738.
- [15] Kowalczyk, A. P., and B. A. Nanes. 2012. "Adherens junction turnover: regulating adhesion through cadherin endocytosis, degradation, and recycling." *Subcell Biochem* 60:197-222. doi: 10.1007/978-94-007-4186-7\_9.
- [16] Nagathihalli, N. S., and N. B. Merchant. 2012. "Src-mediated regulation of E-cadherin and EMT in pancreatic cancer." *Front Biosci (Landmark Ed)* 17:2059-69. doi: 10.2741/4037.
- [17] Park, J. I., S. W. Kim, J. P. Lyons, H. Ji, T. T. Nguyen, K. Cho, M. C. Barton, T. Deroo, K. Vleminckx, R. T. Moon, and P. D. McCrea. 2005. "Kaiso/p120-catenin and TCF/beta-catenin complexes coordinately regulate canonical Wnt gene targets." *Dev Cell* 8 (6):843-54. doi: 10.1016/j.devcel.2005.04.010.
- [18] Bertocchi, C., M. Vaman Rao, and R. Zaidel-Bar. 2012. "Regulation of adherens junction dynamics by phosphorylation switches." *J Signal Transduct* 2012:125295. doi: 10.1155/2012/125295.
- [19] Payan-Carreira, R., M. A. Pires, C. Santos, B. S. Holst, J. Colaço, and H. Rodriguez-Martinez. 2016. "Immunolocalization of E-cadherin and  $\beta$ -catenin in the cyclic and early pregnant canine

- endometrium." *Theriogenology* 86 (4):1092-101. doi: 10.1016/j.theriogenology.2016.03.041.
- [20] Huang, H., S. Wright, J. Zhang, and R. A. Brekken. 2019. "Getting a grip on adhesion: Cadherin switching and collagen signaling." *Biochim Biophys Acta Mol Cell Res* 1866 (11):118472. doi: 10.1016/j.bbamcr.2019.04.002.
- [21] Xiao, X., Y. Ni, C. Yu, L. Li, B. Mao, Y. Yang, D. Zheng, B. Silvestrini, and C. Y. Cheng. 2018. "Src family kinases (SFKs) and cell polarity in the testis." *Semin Cell Dev Biol* 81:46-53. doi: 10.1016/j.semcdb.2017.11.024.
- [22] Xiao, X., Y. Yang, B. Mao, C. Y. Cheng, and Y. Ni. 2019. "Emerging Role for SRC family kinases in junction dynamics during spermatogenesis." *Reproduction*. doi: 10.1530/REP-18-0440.
- [23] Serrels, A., M. Canel, V. G. Brunton, and M. C. Frame. 2011. "Src/FAK-mediated regulation of E-cadherin as a mechanism for controlling collective cell movement: insights from in vivo imaging." *Cell Adh Migr* 5 (4):360-5. doi: 10.4161/cam.5.4.17290.
- [24] Dosch, A. R., X. Dai, A. A. Gaidarski Iii, C. Shi, J. A. Castellanos, M. N. VanSaun, N. B. Merchant, and N. S. Nagathihalli. 2019. "Src kinase inhibition restores E-cadherin expression in dasatinib-sensitive pancreatic cancer cells." *Oncotarget* 10 (10):1056-1069. doi: 10.18632/oncotarget.26621.
- [25] Knights, A. J., A. P. Funnell, M. Crossley, and R. C. Pearson. 2012. "Holding Tight: Cell Junctions and Cancer Spread." *Trends Cancer Res* 8:61-69.
- [26] Bruner, H. C., and P. W. B. Derksen. 2018. "Loss of E-Cadherin-Dependent Cell-Cell Adhesion and the Development and Progression of Cancer." *Cold Spring Harb Perspect Biol* 10 (3). doi: 10.1101/cshperspect.a029330.
- [27] Chaffer, C. L., B. P. San Juan, E. Lim, and R. A. Weinberg. 2016. "EMT, cell plasticity and metastasis." *Cancer Metastasis Rev* 35 (4):645-654. doi: 10.1007/s10555-016-9648-7.
- [28] Nieto, M. A., R. Y. Huang, R. A. Jackson, and J. P. Thiery. 2016. "EMT: 2016." *Cell* 166 (1):21-45. doi: 10.1016/j.cell.2016.06.028.

- [29] Pastushenko, I., A. Brisebarre, A. Sifrim, M. Fioramonti, T. Revenco, S. Boumahdi, A. Van Keymeulen, D. Brown, V. Moers, S. Lemaire, S. De Clercq, E. Minguijon, C. Balsat, Y. Sokolow, C. Dubois, F. De Cock, S. Scozzaro, F. Sopena, A. Lanas, N. D'Haene, I. Salmon, J. C. Marine, T. Voet, P. A. Sotiropoulou, and C. Blanpain. 2018. "Identification of the tumour transition states occurring during EMT." *Nature* 556 (7702):463-468. doi: 10.1038/s41586-018-0040-3.
- [30] Petrova, Y. I., L. Schecterson, and B. M. Gumbiner. 2016. "Roles for E-cadherin cell surface regulation in cancer." *Mol Biol Cell* 27 (21):3233-3244. doi: 10.1091/mbc.E16-01-0058.
- [31] Venhuizen, J. H., F. J. C. Jacobs, P. N. Span, and M. M. Zegers. 2019. "P120 and E-cadherin: Double-edged swords in tumor metastasis." *Seminars in Cancer Biology*. doi: <https://doi.org/10.1016/j.semcancer.2019.07.020>.
- [32] Loh, C. Y., J. Y. Chai, T. F. Tang, W. F. Wong, G. Sethi, M. K. Shanmugam, P. P. Chong, and C. Y. Looi. 2019. "The E-Cadherin and N-Cadherin Switch in Epithelial-to-Mesenchymal Transition: Signaling, Therapeutic Implications, and Challenges." *Cells* 8 (10). doi: 10.3390/cells8101118.
- [33] Ferreira, A. C., G. Suriano, N. Mendes, B. Gomes, X. Wen, F. Carneiro, R. Seruca, and J. C. Machado. 2012. "E-cadherin impairment increases cell survival through Notch-dependent upregulation of Bcl-2." *Hum Mol Genet* 21 (2):334-43. doi: 10.1093/hmg/ddr469.
- [34] Rodriguez, F. J., L. J. Lewis-Tuffin, and P. Z. Anastasiadis. 2012. "E-cadherin's dark side: Possible role in tumor progression." *Biochimica et Biophysica Acta (BBA) - Reviews on Cancer* 1826 (1):23-31. doi: <https://doi.org/10.1016/j.bbcan.2012.03.002>.
- [35] Gallegos, L. L., and J. S. Brugge. 2014. "Live free or die: cell-cell adhesion regulates sensitivity to trail-induced apoptosis." *Dev Cell* 30 (1):3-4. doi: 10.1016/j.devcel.2014.06.031.
- [36] Lu, M., S. Marsters, X. Ye, E. Luis, L. Gonzalez, and A. Ashkenazi. 2014. "E-cadherin couples death receptors to the cytoskeleton to

- regulate apoptosis." *Mol Cell* 54 (6):987-98. doi: 10.1016/j.molcel.2014.04.029.
- [37] Capra, J., and S. Eskelinen. 2017. "Correlation between E-cadherin interactions, survivin expression, and apoptosis in MDCK and ts-Src MDCK cell culture models." *Lab Invest* 97 (12):1453-1470. doi: 10.1038/labinvest.2017.89.
- [38] Carico, E., M. Atlante, E. Giarnieri, S. Raffa, B. Bucci, M. R. Giovagnoli, and A. Vecchione. 2010. "E-cadherin and alpha-catenin expression in normal, hyperplastic and neoplastic endometrium." *Anticancer Res* 30 (12):4993-7.
- [39] Rahnama, F., B. Thompson, M. Steiner, F. Shafiei, P. E. Lobie, and M. D. Mitchell. 2009. "Epigenetic regulation of E-cadherin controls endometrial receptivity." *Endocrinology* 150 (3):1466-1472.
- [40] Horne, A. W., J. O. White, and el-N Lalani. 2002. "Adhesion molecules and the normal endometrium." *BJOG* 109 (6):610-7. doi: 10.1111/j.1471-0528.2002.t01-1-01017.x.
- [41] van der Linden, P. J., A. F. de Goeij, G. A. Dunselman, H. W. Erkens, and J. L. Evers. 1995. "Expression of cadherins and integrins in human endometrium throughout the menstrual cycle." *Fertil Steril* 63 (6):1210-6.
- [42] Fujimoto, J., S. Ichigo, M. Hori, and T. Tamaya. 1996. "Alteration of E-cadherin, alpha- and beta-catenin mRNA expression in human uterine endometrium during the menstrual cycle." *Gynecol Endocrinol* 10 (3):187-91.
- [43] Matsuzaki, S., C. Darcha, E. Maleysson, M. Canis, and G. Mage. 2010. "Impaired down-regulation of E-cadherin and beta-catenin protein expression in endometrial epithelial cells in the mid-secretory endometrium of infertile patients with endometriosis." *J Clin Endocrinol Metab* 95 (7):3437-45. doi: 10.1210/jc.2009-2713.
- [44] Satterfield, M. C., K. A. Dunlap, K. Hayashi, R. C. Burghardt, T. E. Spencer, and F. W. Bazer. 2007. "Tight and adherens junctions in the ovine uterus: differential regulation by pregnancy and progesterone." *Endocrinology* 148 (8):3922-31. doi: 10.1210/en.2007-0321.



- [45] Ryan, P. L., D. L. Baum, J. A. Lenhart, K. M. Ohleth, and C. A. Bagnell. 2001. "Expression of uterine and cervical epithelial cadherin during relaxin-induced growth in pigs." *Reproduction* 122 (6):929-37.
- [46] Guo, B., B. C. Han, Z. Tian, X. M. Zhang, L. X. Jiang, J. X. Liu, and Z. P. Yue. 2009. "Expression and Hormonal Regulation of E-Cadherin in Canine Uterus During Early Pregnancy." *Reprod Domest Anim*. doi: 10.1111/j.1439-0531.2009.01550.x.
- [47] Shih, I. M., M. Y Hsu, R. J. Oldt, M. Herlyn, J. D. Gearhart, and R. J. Kurman. 2002. "The Role of E-cadherin in the Motility and Invasion of Implantation Site Intermediate Trophoblast." *Placenta* 23 (10):706-715. doi: <https://doi.org/10.1053/plac.2002.0864>.
- [48] Kiewisz, J., M. M. Kaczmarek, A. Andronowska, A. Blitek, and A. J. Ziecik. 2011. "Gene expression of WNTs,  $\beta$ -catenin and E-cadherin during the periimplantation period of pregnancy in pigs--involvement of steroid hormones." *Theriogenology* 76 (4):687-99. doi: 10.1016/j.theriogenology.2011.03.022.
- [49] Dudley, J. S., C. R. Murphy, M. B. Thompson, T. Carter, and B. M. McAllan. 2018. "Uterine Epithelial Cells Undergo a Plasma Membrane Transformation During Early Pregnancy in the Domestic Cat (*Felis catus*)." *The Anatomical Record* 301 (9):1497-1505. doi: 10.1002/ar.23895.
- [50] Sakuragi, N., M. Nishiya, K. Ikeda, T. Ohkouch, E. E. Furth, H. Hareyama, C. Satoh, and S. Fujimoto. 1994. "Decreased E-cadherin expression in endometrial carcinoma is associated with tumor dedifferentiation and deep myometrial invasion." *Gynecol Oncol* 53 (2):183-9. doi: 10.1006/gyno.1994.1113.
- [51] Siitonen, S. M., J. T. Kononen, H. J. Helin, I. S. Rantala, K. A. Holli, and J. J. Isola. 1996. "Reduced E-cadherin expression is associated with invasiveness and unfavorable prognosis in breast cancer." *Am J Clin Pathol* 105 (4):394-402. doi: 10.1093/ajcp/105.4.394.
- [52] Koyuncuoglu, M., E. Okyay, B. Saatli, S. Olgan, M. Akin, and U. Saygili. 2012. "Tumor budding and E-Cadherin expression in endometrial carcinoma: are they prognostic factors in endometrial

- cancer?" *Gynecol Oncol* 125 (1):208-13. doi: 10.1016/j.ygyno.2011.12.433.
- [53] Yalta, T., L. Atay, F. Atalay, M. Caydere, M. Gonultas, and H. Ustun. 2009. "E-cadherin expression in endometrial malignancies: comparison between endometrioid and non-endometrioid carcinomas." *J Int Med Res* 37 (1):163-8. doi: 10.1177/147323000903700119.
- [54] Mell, L. K., J. J. Meyer, M. Tretiakova, A. Khramtsov, C. Gong, S. D. Yamada, A. G. Montag, and A. J. Mundt. 2004. "Prognostic significance of E-cadherin protein expression in pathological stage I-III endometrial cancer." *Clin Cancer Res* 10 (16):5546-53. doi: 10.1158/1078-0432.CCR-0943-03.
- [55] González-Rodilla, I., L. Aller, J. Llorca, A. B. Muñoz, V. Verna, J. Estévez, and J. Schneider. 2013. "The E-Cadherin expression vs. tumor cell proliferation paradox in endometrial cancer." *Anticancer Res* 33 (11):5091-5.
- [56] Sugihara, T. 2016. "Loss of Adherens Junction Protein E-Cadherin is a Biomarker of High- Grade Histology and Poor Prognosis in Endometrial Cancer | Insight Medical Publishing." *Annals of Clinical and Laboratory Research* 4. doi: 10.21767/2386-5180.100055.
- [57] Schlosshauer, P. W., L. H. Ellenson, and R. A. Soslow. 2002. "Beta-catenin and E-cadherin expression patterns in high-grade endometrial carcinoma are associated with histological subtype." *Mod Pathol* 15 (10):1032-7. doi: 10.1097/01.MP.0000028573.34289.04.
- [58] Gil da Costa, R. M., M. Santos, I. Amorim, C. Lopes, P. D. Pereira, and A. M. Faustino. 2009. "An immunohistochemical study of feline endometrial adenocarcinoma." *J Comp Pathol* 140 (4):254-9. doi: 10.1016/j.jcpa.2008.12.006.
- [59] Saraiva, A. L., Payan-Carreira, R., Gärtner, F., Pires, M. A. 2012. "Feline Endometrial Adenocarcinomas." In *Adenocarcinoma: Pathogenesis, Treatment and Prognosis*, edited by M. A. Alcalá Longoria, J. I., 175-189. Nova Science Publishers Inc.
- [60] Payan-Carreira, R., A. L. Saraiva, T. Santos, H. Vilhena, A. Sousa, C. Santos, and M. A. Pires. 2013. "Feline Endometrial

- Adenocarcinoma in Females < 1 Year Old: A Description of Four Cases." *Reproduction in Domestic Animals* 48 (5):e70-e77. doi: 10.1111/rda.12190.
- [61] Sontas, B. H., Ö Erdogan, S. Ö Apaydin Enginler, Ö Turna Yilmaz, G. Şennazli, and H. Ekici. 2013. "Endometrial adenocarcinoma in two young queens." *Journal of Small Animal Practice* 54 (3):156-159. doi: 10.1111/j.1748-5827.2012.01307.x.
- [62] Sapierzynski, R. A. Dolka, I. Cywinska, A. 2009. "Multiple pathologies of the feline uterus: a case report." *Veterinarni Medicina* 52 (7):345–350.
- [63] Pires, M. A., A. L. Saraiva, H. Vilhena, S. Miranda, I. Fonseca, P. Moreira, A. M. Alves, R. Paiva, and R. Payan-Carreira. 2013. "Endometrial adenocarcinoma in a cat with abdominal metastasis." *Journal of Comparative Pathology* 148 (1):67-67.
- [64] Saraiva, A. L., R. Payan-Carreira, F. Gärtner, M. R. Fortuna da Cunha, A. Rêma, F. Faria, L. M. Lourenço, and M.os A Pires. 2015. "An immunohistochemical study on the expression of sex steroid receptors, Ki-67 and cytokeratins 7 and 20 in feline endometrial adenocarcinomas." *BMC Vet Res* 11:204. doi: 10.1186/s12917-015-0530-6.
- [65] Pires, M. A., F. Seixas, C. Palmeira, and R. Payan-Carreira. 2010. "Histopathologic and Immunohistochemical Exam in One Case of Canine Endometrial Adenocarcinoma." *Reproduction in Domestic Animals* 45 (3):545-549. doi: 10.1111/j.1439-0531.2008.01243.x.
- [66] Restucci, B., S. Papparella, G. De Vico, and P. Maiolino. 1997. "E cadherin expression in normal and neoplastic canine mammary gland." *Journal of Comparative Pathology* 116 (2):191-202. doi: [https://doi.org/10.1016/S0021-9975\(97\)80076-X](https://doi.org/10.1016/S0021-9975(97)80076-X).
- [67] Sarli, G., R. Preziosi, L. De Tolla, B. Brunetti, and C. Benazzi. 2004. "E-cadherin immunoreactivity in canine mammary tumors." *J Vet Diagn Invest* 16 (6):542-7. doi: 10.1177/104063870401600608.
- [68] Gama, A., J. Paredes, F. Gartner, A. Alves, and F. Schmitt. 2008. "Expression of E-cadherin, P-cadherin and beta-catenin in canine malignant mammary tumours in relation to clinicopathological

- parameters, proliferation and survival." *Vet J* 177 (1):45-53. doi: 10.1016/j.tvjl.2007.05.024.
- [69] Figueira, A. C., C. Gomes, J. Tavares de Oliveira, H. Vilhena, J. Carvalheira, A. J. F. de Matos, P. Dias Pereira, and F. Gaertner. 2014. "Aberrant P-cadherin expression is associated to aggressive feline mammary carcinomas." *Bmc Veterinary Research* 10. doi: 10.1186/s12917-014-0270-z.
- [70] Brunetti, B., G. Sarli, R. Preziosi, S. Leprotti, and C. Benazzi. 2003. "E-cadherin expression in canine mammary carcinomas with regional lymph node metastases." *J Vet Med A Physiol Pathol Clin Med* 50 (10):496-500. doi: 10.1111/j.1439-0442.2003.00577.x.
- [71] De Matos, A. J., C. C. Lopes, A. M. Faustino, J. G. Carvalheira, G. R. Rutteman, and M.e F Gärtner. 2007. "E-cadherin, beta-catenin, invasion and lymph node metastases in canine malignant mammary tumours." *APMIS* 115 (4):327-34. doi: 10.1111/j.1600-0463.2007.apm\_544.x.
- [72] Park, D., R. KÅResen, U. Axcrona, T. Noren, and T. Sauer. 2007. "Expression pattern of adhesion molecules (E-cadherin,  $\alpha$ -,  $\beta$ -,  $\gamma$ -catenin and claudin-7), their influence on survival in primary breast carcinoma, and their corresponding axillary lymph node metastasis." *APMIS* 115 (1):52-65. doi: 10.1111/j.1600-0463.2007.apm\_524.x.
- [73] Figueira, A. C. Gomes, C. Vilhena, H. Miranda, S. Carvalheira, J De Matos, A. J. Dias-Pereira, P. Gärtner F. 2015. "Characterization of alpha-, beta- and p120-Catenin Expression in Feline Mammary Tissues and their Relation with E- and P-Cadherin." *Anticancer Res* 35 (6):3361-3369.
- [74] Palacios, J., N. Benito, A. Pizarro, A. Suarez, J. Espada, A. Cano, and C. Gamallo. 1995. "Anomalous expression of P-cadherin in breast carcinoma. Correlation with E-cadherin expression and pathological features." *Am J Pathol* 146 (3):605-612.
- [75] Gama, A., J. Paredes, M. F. Milanezi, J. S. Reis-Filho, F. Gartner, and F. C. Schmitt. 2002. "P-cadherin expression in canine lactating mammary gland." *J Cell Biochem* 86 (3):420-421. doi: 10.1002/jcb.10245.

- [76] Gama, A., J. Paredes, A. Albergaria, F. Gartner, and F. Schmitt. 2004. "P-cadherin expression in canine mammary tissues." *J Comp Pathol* 130 (1):13-20. doi: 10.1016/s0021-9975(03)00064-1.
- [77] Visvader, J. E., and J. Stingl. 2014. "Mammary stem cells and the differentiation hierarchy: current status and perspectives." *Genes Dev* 28 (11):1143-1158. doi: 10.1101/gad.242511.114.
- [78] Derksen, P. W., T. M. Braumuller, E. van der Burg, M. Hornsveld, E. Mesman, J. Wesseling, P. Krimpenfort, and J. Jonkers. 2011. "Mammary-specific inactivation of E-cadherin and p53 impairs functional gland development and leads to pleomorphic invasive lobular carcinoma in mice." *Dis Model Mech* 4 (3):347-358. doi: 10.1242/dmm.006395.
- [79] Xi, Y., X. Zhang, Z. Yang, Q. Guo, Z. Zhang, S. Chen, H. Zheng, and B. Hua. 2019. "Prognostic significance of P-cadherin expression in breast cancer: Protocol for a meta-analysis." *Medicine (Baltimore)* 98 (12):e14924. doi: 10.1097/MD.00000000000014924.
- [80] Heimann, R., F. Lan, R. McBride, and S. Hellman. 2000. "Separating favorable from unfavorable prognostic markers in breast cancer: the role of E-cadherin." *Cancer Res* 60 (2):298-304.
- [81] Siegel, R. L., K. D. Miller, and A. Jemal. 2018. "Cancer statistics, 2018." *CA Cancer J Clin* 68 (1):7-30. doi: 10.3322/caac.21442.
- [82] Misdorp, W. 2002. "Tumors of the Mammary Gland." *Tumors in Domestic Animals*:575-606. doi: doi:10.1002/9780470376928.ch12.
- [83] Gama, A., and F. Schmitt. 2012. "Cadherin cell adhesion system in canine mammary cancer: a review." *Vet Med Int* 2012:357187. doi: 10.1155/2012/357187.
- [84] Rutteman, G. R., S. J. Withrow, and E. G. MacEwen. 2001. "Tumours of the mammary gland." In *Small Animal Clinical Oncology*, edited by SJ Withrow and EG MacEwen, 455-477. Philadelphia: W. B. Saunders Comp.
- [85] Millanta, F., G. Lazzeri, I. Vannozzi, P. Viacava, and A. Poli. 2002. "Correlation of vascular endothelial growth factor expression to overall survival in feline invasive mammary carcinomas." *Vet Pathol* 39 (6):690-696. doi: 10.1354/vp.39-6-690.

- [86] Seixas, F., C. Palmeira, M. A. Pires, M. J. Bento, and C. Lopes. 2011. "Grade is an independent prognostic factor for feline mammary carcinomas: a clinicopathological and survival analysis." *Vet J* 187 (1):65-71. doi: 10.1016/j.tvjl.2009.10.030.
- [87] Lombaerts, M., T. van Wezel, K. Philippo, J. W. Dierssen, R. M. Zimmerman, J. Oosting, R. van Eijk, P. H. Eilers, B. van de Water, C. J. Cornelisse, and A. M. Cleton-Jansen. 2006. "E-cadherin transcriptional downregulation by promoter methylation but not mutation is related to epithelial-to-mesenchymal transition in breast cancer cell lines." *Br J Cancer* 94 (5):661-71. doi: 10.1038/sj.bjc.6602996.
- [88] Horne, H. N., H. Oh, M. E. Sherman, M. Palakal, S. M. Hewitt, M. K. Schmidt, R. L. Milne, D. Hardisson, J. Benitez, C. Blomqvist, M. K. Bolla, H. Brenner, J. Chang-Claude, R. Cora, F. J. Couch, K. Cuk, P. Devilee, D. F. Easton, D. M. Eccles, U. Eilber, J. M. Hartikainen, P. Heikkilä, B. Holleczeck, M. J. Hooning, M. Jones, R. Keeman, A. Mannermaa, J. W. M. Martens, T. A. Muranen, H. Nevanlinna, J. E. Olson, N. Orr, J. I. A. Perez, P. D. P. Pharoah, K. J. Ruddy, K. U. Saum, M. J. Schoemaker, C. Seynaeve, R. Sironen, V. T. H. BM Smit, A. J. Swerdlow, M. Tengström, A. S. Thomas, A. M. Timmermans, R. A. E.M Tollenaar, M. A. Troester, C. J. van Asperen, C. H. M. van Deurzen, F. F. Van Leeuwen, L. J. Van't Veer, M. García-Closas, and J. D. Figueroa. 2018. "E-cadherin breast tumor expression, risk factors and survival: Pooled analysis of 5,933 cases from 12 studies in the Breast Cancer Association Consortium." *Sci Rep* 8 (1):6574. doi: 10.1038/s41598-018-23733-4.
- [89] Borcharding, N., K. Cole, P. Kluz, M. Jorgensen, R. Kolb, A. Bellizzi, and W. Zhang. 2018. "Re-Evaluating E-Cadherin and  $\beta$ -Catenin: A Pan-Cancer Proteomic Approach with an Emphasis on Breast Cancer." *Am J Pathol* 188 (8):1910-1920. doi: 10.1016/j.ajpath.2018.05.003.
- [90] Ye, Y. Tellez, J. D. Durazo, M. Belcher, M. Yearsley, K. Barsky, S. H. 2010. "E-Cadherin Accumulation within the Lymphovascular

- Embolus of Inflammatory Breast Cancer Is Due to Altered Trafficking." *Anticancer Research* 30:3903-3910.
- [91] Gamallo, C., J. Palacios, A. Suarez, A. Pizarro, P. Navarro, M. Quintanilla, and A. Cano. 1993. "Correlation of E-cadherin expression with differentiation grade and histological type in breast carcinoma." *Am J Pathol* 142 (4):987-93.
- [92] Kovács, A., J. Dhillon, and R. A. Walker. 2003. "Expression of P-cadherin, but not E-cadherin or N-cadherin, relates to pathological and functional differentiation of breast carcinomas." *Mol Pathol* 56 (6):318-22. doi: 10.1136/mp.56.6.318.
- [93] Li, Z., S. Yin, L. Zhang, W. Liu, and B. Chen. 2017. "Prognostic value of reduced E-cadherin expression in breast cancer: a meta-analysis." *Oncotarget* 8 (10):16445-16455. doi: 10.18632/oncotarget.14860.
- [94] Rakha, E. A., D. Abd El Rehim, S. E. Pinder, S. A. Lewis, and I. O. Ellis. 2005. "E-cadherin expression in invasive non-lobular carcinoma of the breast and its prognostic significance." *Histopathology* 46 (6):685-93. doi: 10.1111/j.1365-2559.2005.02156.x.
- [95] Charafe-Jauffret, E., C. Tarpin, V. J. Bardou, F. Bertucci, C. Ginestier, A. C. Braud, B. Puig, J. Geneix, J. Hassoun, D. Birnbaum, J. Jacquemier, and P. Viens. 2004. "Immunophenotypic analysis of inflammatory breast cancers: identification of an 'inflammatory signature'." *J Pathol* 202 (3):265-73. doi: 10.1002/path.1515.
- [96] Gould Rothberg, B. E., and M. B. Bracken. 2006. "E-cadherin immunohistochemical expression as a prognostic factor in infiltrating ductal carcinoma of the breast: a systematic review and meta-analysis." *Breast Cancer Res Treat* 100 (2):139-48. doi: 10.1007/s10549-006-9248-2.
- [97] Gillett, C. E., D. W. Miles, K. Ryder, D. Skilton, R. D. Liebman, R. J. Springall, D. M. Barnes, and A. M. Hanby. 2001. "Retention of the expression of E-cadherin and catenins is associated with shorter survival in grade III ductal carcinoma of the breast." *J Pathol* 193 (4):433-41. doi: 10.1002/path.831.

- [98] Wang, D., L. Su, D. Huang, H. Zhang, D. M. Shin, and Z. G. Chen. 2011. "Downregulation of E-Cadherin enhances proliferation of head and neck cancer through transcriptional regulation of EGFR." *Mol Cancer* 10:116. doi: 10.1186/1476-4598-10-116.
- [99] Reis, A. L., J. Carvalheira, F. C. Schmitt, and F. Gartner. 2003. "Immunohistochemical study of the expression of E-cadherin in canine mammary tumours." *Vet Rec* 152 (20):621-624. doi: 10.1136/vr.152.20.621.
- [100] Brunetti, B., G. Sarli, R. Preziosi, I. Monari, and C. Benazzi. 2005. "E-cadherin and beta-catenin reduction influence invasion but not proliferation and survival in canine malignant mammary tumors." *Vet Pathol* 42 (6):781-787. doi: 10.1354/vp.42-6-781.
- [101] Torres, L. N., J. M. Matera, C. H. Vasconcellos, J. L. Avanzo, F. J. Hernandez-Blazquez, and M. L. Dagli. 2005. "Expression of connexins 26 and 43 in canine hyperplastic and neoplastic mammary glands." *Vet Pathol* 42 (5):633-641. doi: 10.1354/vp.42-5-633.
- [102] Nowak, M., J. A. Madej, and P. Dziegiel. 2007. "Expression of E-cadherin, beta-catenin and Ki-67 antigen and their reciprocal relationships in mammary adenocarcinomas in bitches." *Folia Histochem Cytobiol* 45 (3):233-238.
- [103] Nowak, M., J. A. Madej, M. Podhorska-Okolow, and P. Dziegiel. 2008. "Expression of extracellular matrix metalloproteinase (MMP-9), E-cadherin and proliferation-associated antigen Ki-67 and their reciprocal correlation in canine mammary adenocarcinomas." *In Vivo* 22 (4):463-469.
- [104] Rodo, A., and E. Malicka. 2008. "E-cadherin immunohistochemical expression in mammary gland neoplasms in bitches." *Pol J Vet Sci* 11 (1):47-54.
- [105] Yoshida, K., S. Yoshida, N. Choisunirachon, T. Saito, K. Matsumoto, K. Saeki, M. Mochizuki, R. Nishimura, N. Sasaki, and T. Nakagawa. 2014. "The relationship between clinicopathological features and expression of epithelial and mesenchymal markers in spontaneous canine mammary gland tumors." *J Vet Med Sci* 76 (10):1321-1327. doi: 10.1292/jvms.14-0104.



- [106] Dias-Pereira, P., and F. Gärtner. 2003. "Expression of E-cadherin in normal, hyperplastic and neoplastic feline mammary tissue." *Vet Rec* 153 (10):297-302.
- [107] Takauji, S. R., M. Watanabe, R. Uyama, T. Nakagawa, N. Miyajima, M. Mochizuki, R. Nishimura, S. Sugano, and N. Sasaki. 2007. "Expression and subcellular localization of E-cadherin, alpha-catenin, and beta-catenin in 8 feline mammary tumor cell lines." *J Vet Med Sci* 69 (8):831-834. doi: 10.1292/jvms.69.831.
- [108] Penafiel-Verdu, C., A. J. Buendia, J. A. Navarro, G. A. Ramirez, M. Vilafranca, J. Altimira, and J. Sanchez. 2012. "Reduced expression of E-cadherin and beta-catenin and high expression of basal cytokeratins in feline mammary carcinomas with regional metastasis." *Vet Pathol* 49 (6):979-987. doi: 10.1177/0300985812436744.
- [109] Zappulli, V., S. De Cecco, D. Trez, D. Caliari, L. Aresu, and M. Castagnaro. 2012. "Immunohistochemical expression of E-cadherin and beta-catenin in feline mammary tumours." *J Comp Pathol* 147 (2-3):161-170. doi: 10.1016/j.jcpa.2012.02.004.
- [110] Howard, E. M., S. K. Lau, R. H. Lyles, G. G. Birdsong, J. N. Umbreit, and R. Kochhar. 2005. "Expression of e-cadherin in high-risk breast cancer." *J Cancer Res Clin Oncol* 131 (1):14-8. doi: 10.1007/s00432-004-0618-z.
- [111] Kleer, C. G., K. L. van Golen, T. Braun, and S. D. Merajver. 2001. "Persistent E-cadherin expression in inflammatory breast cancer." *Mod Pathol* 14 (5):458-64. doi: 10.1038/modpathol.3880334.
- [112] Chu, K., K. M. Boley, R. Moraes, S. H. Barsky, and F. M. Robertson. 2013. "The paradox of E-cadherin: role in response to hypoxia in the tumor microenvironment and regulation of energy metabolism." *Oncotarget* 4 (3):446-462. doi: 10.18632/oncotarget.872.
- [113] Sousa, B., J. Pereira, and J. Paredes. 2019. "The Crosstalk Between Cell Adhesion and Cancer Metabolism." *Int J Mol Sci* 20 (8). doi: 10.3390/ijms20081933.
- [114] Yoshida, R. Kimura, N. Harada, Y. Ohuchi, N. 2001. "The loss of E-cadherin,  $\alpha$ - and  $\beta$ -catenin expression is associated with metastasis

- and poor prognosis in invasive breast cancer." *International Journal of Oncology* 18 (13):513-520. doi: 10.3892/ijo.18.3.513.
- [115] Yang, L., X. W. Wang, L. P. Zhu, H. L. Wang, B. Wang, Q. Zhao, and X. Y. Wang. 2018. "Significance and prognosis of epithelial-cadherin expression in invasive breast carcinoma." In *Oncol Lett*, 1659-1665.
- [116] Bukholm, I. K., J. M. Nesland, and A. L. Børresen-Dale. 2000. "Re-expression of E-cadherin,  $\alpha$ -catenin and  $\beta$ -catenin, but not of  $\gamma$ -catenin, in metastatic tissue from breast cancer patients." *The Journal of Pathology* 190 (1):15-19. doi: 10.1002/(SICI)1096-9896(200001)190:1<15::AID-PATH489>3.0.CO;2-L.
- [117] Elisha, Y., V. Kalchenko, Y. Kuznetsov, and B. Geiger. 2018. "Dual role of E-cadherin in the regulation of invasive collective migration of mammary carcinoma cells." *Sci Rep* 8 (1):4986. doi: 10.1038/s41598-018-22940-3.
- [118] Kalluri, R., and R. A. Weinberg. 2009. "The basics of epithelial-mesenchymal transition." *J Clin Invest* 119 (6):1420-1428. doi: 10.1172/jci39104.
- [119] Thiery, J. P., H. Acloque, R. Y. Huang, and M. A. Nieto. 2009. "Epithelial-mesenchymal transitions in development and disease." *Cell* 139 (5):871-890. doi: 10.1016/j.cell.2009.11.007.
- [120] Berx, G., A. M. Cleton-Jansen, K. Strumane, W. J. de Leeuw, F. Nollet, F. van Roy, and C. Cornelisse. 1996. "E-cadherin is inactivated in a majority of invasive human lobular breast cancers by truncation mutations throughout its extracellular domain." *Oncogene* 13 (9):1919-1925.
- [121] De Leeuw, W. J., G. Berx, C. B. Vos, J. L. Peterse, M. J. Van de Vijver, S. Litvinov, F. Van Roy, C. J. Cornelisse, and A. M. Cleton-Jansen. 1997. "Simultaneous loss of E-cadherin and catenins in invasive lobular breast cancer and lobular carcinoma in situ." *J Pathol* 183 (4):404-411. doi: 10.1002/(sici)1096-9896(199712)183:4<404::aid-path1148>3.0.co;2-9.
- [122] Sarrio, D., G. Moreno-Bueno, D. Hardisson, C. Sanchez-Estevéz, M. Guo, J. G. Herman, C. Gamallo, M. Esteller, and J. Palacios. 2003.

- "Epigenetic and genetic alterations of APC and CDH1 genes in lobular breast cancer: relationships with abnormal E-cadherin and catenin expression and microsatellite instability." *Int J Cancer* 106 (2):208-215. doi: 10.1002/ijc.11197.
- [123] Tsuji, T., S. Ibaragi, and G. F. Hu. 2009. "Epithelial-mesenchymal transition and cell cooperativity in metastasis." *Cancer Res* 69 (18):7135-7139. doi: 10.1158/0008-5472.can-09-1618.
- [124] Drasin, D. J., T. P. Robin, and H. L. Ford. 2011. "Breast cancer epithelial-to-mesenchymal transition: examining the functional consequences of plasticity." *Breast Cancer Res* 13 (6):226. doi: 10.1186/bcr3037.
- [125] Raposo-Ferreira, T. M. M., B. K. Brisson, A. C. Durham, R. Laufer-Amorim, V. Kristiansen, E. Pure, S. W. Volk, and K. Sorenmo. 2018. "Characteristics of the Epithelial-Mesenchymal Transition in Primary and Paired Metastatic Canine Mammary Carcinomas." *Vet Pathol* 55 (5):622-633. doi: 10.1177/0300985818776054.
- [126] Im, K. S., J. H. Kim, N. H. Kim, C. H. Yu, T. Y. Hur, and J. H. Sur. 2012. "Possible role of Snail expression as a prognostic factor in canine mammary neoplasia." *J Comp Pathol* 147 (2-3):121-128. doi: 10.1016/j.jcpa.2011.12.002.
- [127] Gamba, C. O., M. A. Rodrigues, D. A. Gomes, A. Estrela-Lima, E. Ferreira, and G. D. Cassali. 2015. "The Relationship Between E-Cadherin and its Transcriptional Repressors in Spontaneously Arising Canine Invasive Micropapillary Mammary Carcinoma." *J Comp Pathol* 153 (4):256-265. doi: 10.1016/j.jcpa.2015.08.006.
- [128] Gamba, C. O., K. A. Damasceno, I. C. Ferreira, M. A. Rodrigues, D. A. Gomes, M. R. Alves, R. M. Rocha, A. E. Lima, E. Ferreira, and G. D. Cassali. 2019. "The investigation of transcriptional repression mediated by ZEB2 in canine invasive micropapillary carcinoma in mammary gland." *PLoS One* 14 (1):e0209497. doi: 10.1371/journal.pone.0209497.
- [129] Pang, L. Y., A. Cervantes-Arias, R. W. Else, and D. J. Argyle. 2011. "Canine Mammary Cancer Stem Cells are Radio- and Chemo-Resistant and Exhibit an Epithelial-Mesenchymal Transition

- Phenotype." *Cancers (Basel)* 3 (2):1744-1762. doi: 10.3390/cancers3021744.
- [130] Hollestelle, A., J. K. Peeters, M. Smid, M. Timmermans, L. C. Verhoog, P. J. Westenend, A. A. Heine, A. Chan, A. M. Sieuwerts, E. A. Wiemer, J. G. Klijn, P. J. van der Spek, J. A. Foekens, M. Schutte, M. A. den Bakker, and J. W. Martens. 2013. "Loss of E-cadherin is not a necessity for epithelial to mesenchymal transition in human breast cancer." *Breast Cancer Res Treat* 138 (1):47-57. doi: 10.1007/s10549-013-2415-3.
- [131] Timmermans-Sprang, E., R. Collin, A. Henkes, M. Philipsen, and J. A. Mol. 2019. "P-cadherin mutations are associated with high basal Wnt activity and stemness in canine mammary tumor cell lines." *Oncotarget* 10 (31):2930-2946. doi: 10.18632/oncotarget.26873.
- [132] Paredes, J., J. Figueiredo, A. Albergaria, P. Oliveira, J. Carvalho, A. S. Ribeiro, J. Caldeira, A. M. Costa, J. Simoes-Correia, M. J. Oliveira, H. Pinheiro, S. S. Pinho, R. Mateus, C. A. Reis, M. Leite, M. S. Fernandes, F. Schmitt, F. Carneiro, C. Figueiredo, C. Oliveira, and R. Seruca. 2012. "Epithelial E- and P-cadherins: role and clinical significance in cancer." *Biochim Biophys Acta* 1826 (2):297-311. doi: 10.1016/j.bbcan.2012.05.002.
- [133] Buendia, A. J., C. Penafiel-Verdu, J. A. Navarro, M. Vilafranca, and J. Sanchez. 2014. "N-cadherin expression in feline mammary tumors is associated with a reduced E-cadherin expression and the presence of regional metastasis." *Vet Pathol* 51 (4):755-758. doi: 10.1177/0300985813505115.
- [134] Figueira, A. C., C. Gomes, N. Mendes, I. F. Amorim, A. J. F. Matos, P. Dias-Pereira, and F. Gärtner. 2016. "An in vitro and in vivo characterization of the cadherin-catenin adhesion complex in a feline mammary carcinoma cell line." *Clin Diagn Pathol* 1 (1):1-8. doi: 10.15761/NRD.1000103.